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(54) Title: **NOVEL GLUCANS AND NOVEL GLUCANSUCRASES DERIVED FROM LACTIC ACID BACTERIA**

(57) Abstract: The invention pertains to novel glucans capable of being produced by glucosyltransferase activity of a lactic acid bacterium on a sucrose substrate, the glucan having an average molecular weight between 10 kDa and 1 GDa, consisting essentially of $\alpha(1,3)$ - and $\alpha(1,6)$ -linked anhydroglucose units (AGU) and to glucansucrases capable of producing these glucans from sucrose. The glucans have thickening and anti-corrosive properties. The glucans can be chemically modified.

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Novel glucans and novel glucansucrases derived from lactic acid bacteria

[0001] The present invention is in the field of enzymatic production of biomolecules. The invention is particularly concerned with novel glucans derived from lactic acid bacteria, with novel glucosyl-transferases derived from such bacteria and with a process for
5 production of new and useful glucans from sucrose.

Background of the invention

[0002] Several bacteria are known to produce exopolysaccharides, i.e. polysaccharides secreted into the culture medium. Well-known examples of bacterial exopolysaccharides include xanthan from *Xanthomonas campestris*, gellan from *Sphingomonas paucimobilis*
10 and pullulan from *Aureobasidium pullulans*. Lactic acid bacteria known to produce exopolysaccharides include *Leuconostoc mesenteroides* strains producing dextrans, $\alpha(1\rightarrow 6)$ -linked poly-anhydroglucose, and alternans i.e. poly-anhydroglucoses having alternating $\alpha(1\rightarrow 6)$ and $\alpha(1\rightarrow 3)$ -linkages, oral *Streptococcus* strains producing glucans responsible for dental plaque formation, and a particular *Lactobacillus reuteri* strain producing
15 $\alpha(1,6)$ - and $\alpha(1,4)$ -linked anhydroglucose (Van Geel-Schutten, *et al.*, *Appl. Environ. Microbiol.* (1999) 65, 3008-3014). The properties of exopolysaccharides depend on the type of monosaccharide units, the type of linkages, the degree and type of branching, the length of the polysaccharide chain, the molecular weight and the conformation of the polymers.

[0003] Argüello-Morales *et al.* (*FEMS Microbiol. Lett.* 182 (2000) 81-85) describe an alternansucrase from *Leuconostoc mesenteroides* NRRL B-1355. Monchois *et al.* (*Gene* 182 (1996) 23-32; *FEMS Microbiol. Lett.* 159 (1998) 307-315) for instance describe two different dextransucrases from *Lc. mesenteroides* NRRL B-1299. A method for selecting *Leuconostoc mesenteroides* strains that produce a high proportion of alternan to dextran is
25 described in US 5,789,209. The prior art does not disclose or suggest other lactic acid bacteria than *Leuconostoc* or *Streptococcus* that are capable of producing glucans having both $\alpha(1\rightarrow 6)$ and $\alpha(1\rightarrow 3)$ -linkages.

Summary of the invention

[0004] Several lactic acid bacteria strains were found, according to the invention, to be
30 capable of producing a particular class of glucans. These glucans have in common that their anhydroglucose units (AGU) are linked $\alpha(1,3)$ - and/or $\alpha(1,6)$ -glucosidic bonds, i.e. they are α -glucans largely or completely devoid of $\alpha(1,4)$ -bonds. These glucans may be of

the alternan (alternating $\alpha(1,3)$ and $\alpha(1,6)$ linkages), mutan (mixed $\alpha(1,3)$ and $\alpha(1,6)$ linkages, usually $\alpha(1,3)$ predominant) or dextran (mainly $\alpha(1,6)$ linkages, some $\alpha(1,3)$) type, or other type. The glucans can be produced from sucrose, using sucrase enzymes which are active in the lactic acid bacteria. They can be produced on a large scale and isolated in a commercially feasible way, as the glucans are produced outside the bacterial cell, or even in the absence of the bacteria, using isolated sucrase enzymes. The glucans are produced by food-grade strains and have interesting properties, such as prebiotic utility or thickening of water-based compositions.

[0005] The invention is concerned with these novel glucans, with the lactic acid bacterial, especially *Lactobacillus* strains and their enzymic proteins that produce these glucans from sucrose, as well as with methods for producing the glucans using the strains and/or their enzymes, with nucleotide sequences encoding these enzymic proteins which convert sucrose, with the use of the glucans as thickeners, prebiotics, anticorrosives, etc., and as starting materials for modified glucans.

15 *Description of the invention*

[0006] The invention pertains to *Lactobacillus* strains containing a glucosyltransferase (glucansucrase) capable of producing a glucan having at least 10 anhydroglucose units (AGU) having a backbone consisting essentially of $\alpha(1,3)$ - and/or $\alpha(1,6)$ -linked AGU, in the presence of sucrose. Such strains can be found among current sources of *Lactobacilli*, such as food sources, silage, mammalian samples etc. These strains containing the glucosyltransferases and producing the glucans can be identified by isolating *Lactobacillus* strains from these sources, growing them on sucrose and analysing the polysaccharide product using suitable analytical methods such as chromatography. The genes encoding these glucosyltransferases can be identified by amplifying nucleotide sequence fragments of the strain using primers based on known glucosyltransferase genes and retaining the positive strains (see examples). Several glucan-producing strains were isolated and identified from different sources and different *Lactobacillus* species, such as *Lb. reuteri*, *Lb. fermentum*, *Lb. sake* and *Lb. parabuechneri* or related species. The glucosyltransferases from these glucan-producing strains were also identified and, completely or partly, sequenced (see Examples).

[0007] The novel glucans of the invention are capable of being produced by glucosyltransferase (glucansucrase) activity of a lactic acid bacterium on a sucrose donor substrate. The glucans have an average molecular weight between 10 kDa and 1 GDa, and

consist essentially of $\alpha(1,3)$ - and/or $\alpha(1,6)$ -linked anhydroglucose units (AGU), to which side-chains also consisting of $\alpha(1,3)$ - and/or $\alpha(1,6)$ -linked AGU may be attached.

[0008] In particular, the glucans according to the invention either comprise 15-80% of $\alpha(1,3)$ -linked AGU, 2-80%, especially 4-80% and more especially 15-80% of $\alpha(1,6)$ -
5 linked and 2-25% of $\alpha(1,3,6)$ -linked (branching) AGU, or 80-99% of $\alpha(1,6)$ -linked AGU and 1-20% of $\alpha(1,3)$ -linked or $\alpha(1,3,6)$ -linked (branching) AGU, in particular 1-15% of $\alpha(1,3)$ -linked AGU and 5-15% of $\alpha(1,3)$ - and $\alpha(1,3,6)$ -linked units taken together. Thus, the invention covers a glucan having an average molecular weight of 50 kDa to 1 MDa and comprising 25-50%, especially 29-39% of $\alpha(1,3)$ -linked AGU, 20-45%, especially
10 30-40% of $\alpha(1,6)$ -linked AGU, 5-25%, especially 3-13% of $\alpha(1,3,6)$ -linked AGU and 6-30% of terminal AGU. Furthermore, the invention pertains to a glucan having an average molecular weight of 10-50 MDa and comprising 15-26% $\alpha(1,3)$ -linked AGU, 30-50% of $\alpha(1,6)$ -linked AGU, 5-20% of $\alpha(1,3,6)$ -linked AGU and 5-35% of terminal AGU. Also, in another embodiment the invention covers a glucan having an average molecular weight
15 of 1-50 MDa and comprising 40-60% of $\alpha(1,3)$ -linked AGU, 2-20%, especially 2-12% of $\alpha(1,6)$ -linked AGU, 10-25% of $\alpha(1,3,6)$ -linked AGU and 10-30% of terminal AGU. In yet another embodiment, the invention comprises a glucan having an average molecular weight of 10-50 MDa and comprising 80-99%, especially 88-99% and more especially 90-99% of $\alpha(1,6)$ -linked AGU, or 80-90% of $\alpha(1,6)$ - and 1-10% of $\alpha(1,3)$ -linked AGU,
20 the remainder being 1,3,6 linked and terminal AGU.

[0009] The invention also concerns the enzymes originating from lactic acid bacteria, or from recombinant sources, capable of producing the glucans described above starting from sucrose. The enzymes are new and they can be classified as glucansucrases or glucosyltransferases. Their partial sequence information is given below in SEQ ID No's
25 1-10. More complete sequence information is given in SEQ ID No's 11-22. Proteins according to the invention comprise an amino acid sequence exhibiting at least 70%, preferably at least 80%, most preferably at least 90%, amino acid identity with any one of the amino acid sequences of SEQ ID No. 2, 4, 8, 10, 12, 14, 16, 18, 20 and 22 or of stretches of at least 221-224 amino acids thereof, or at least 100 contiguous amino acids
30 exhibiting at least 80%, preferably at least 90%, amino acid identity with these sequences. Further preferred sequences are indicated in the description of the alignment figure given below.

[0010] The enzymes can be used as such for producing the glucans described above, or for producing oligosaccharides and polysaccharides having a similar $\alpha(1,3)$ and/or $\alpha(1,6)$ linked glucan structure. Their genes can also be incorporated in suitable host organisms, to produce alternative glucan-production systems. The invention also pertains to such
5 recombinant, preferably food-grade microorganisms, e.g. bacteria, especially lactic acid bacteria, yeasts, fungi etc., containing the genes of the glucansucrases described above and being capable of expressing the glucansucrases.

[0011] The invention also pertains to a process of producing a glucan as described above. This glucan can be produced by a *Lactobacillus* strain as described above, or by a
10 recombinant micro-organism expressing the glucosyltransferase according to the invention or by an isolated glucosyltransferase according to the invention and a suitable glucose source such as for instance sucrose. The glucosyltransferase may be isolated by conventional means from the culture of a glucosyltransferase-positive lactic acid bacterium, especially a *Lactobacillus* species, or from a recombinant organism expressing
15 the glucosyltransferase gene.

[0012] The glucan and the gluco-oligosaccharides produced by the *Lactobacillus* strains can be recovered from the culture supernatant of *Lactobacillus* strains described above, containing the glucosyltransferase according to the invention. The glucan can comprise at
20 least 20, up to about 100,000 α -anhydroglucose units with the unique structure described above.

[0013] The glucan-producing enzymes according to invention, or at least the most preferred ones, are constitutive in the *Lactobacillus* strains, in that they are always present. This is contrast to most glucan (dextran-) producing *Leuconostoc* strains of the prior art, wherein the enzymes are only expressed upon growth in the presence of sucrose.
25 This allows a more efficient production of glucans by the microorganisms of the invention.

[0014] The glucans according to invention have a variety of useful properties. They are suitable as prebiotics, and thus they can be incorporated in nutritional or pharmaceutical compositions intended for improving the condition of the gastrointestinal tract. For this
30 purpose, they can be used as such or in the form of their oligosaccharides. They can also be combined with other poly- or oligosaccharides, such as fructans, galactans, xylans, arabinans, mannans, indigestible glucans and hetero-oligosaccharides, or with probiotic micro-organisms, including the lactic acid bacteria from which the glucans originate, resulting in synbiotic compositions. The glucans and their shortened homologues are also

useful as bioactive agents, e.g. as immunomodulators, anti-ulcer agents and cholesterol-lowering agents.

[0015] The glucans are also useful as thickening agents. As such they can be incorporated in foodstuffs such as beverages, sauces, dressings, dairy products, in amounts of from 1 g/l to about 100 g/l, especially about 10 to 50 g/l.

[0016] The glucans of the invention are furthermore useful as anticorrosion agents, for example for the protection of ship hulls. For that purpose, they may be applied in the form of solutions or suspensions, by spraying, coating, dipping and other techniques known in the art of corrosion control.

[0017] The glucans can be used as such. They can also be modified by physical or chemical means. Suitable examples of chemical modification include oxidation, especially 2,3- or 3,4-oxidation using periodate or hypohalite, in glucans having α -1,6 linkages, or 6-oxidation using nitroxyls with peracid or hypohalite in glucans having α -1,3 linkages. Hypohalite oxidation resulting in ring-opened 2,3- or 3,4-dicarboxy-anhydroglucose units (see e.g. EP-A-427349), while periodate oxidation results in ring-opened 2,3- or 3,4-dialdehyde-anhydroglucose units (see e.g. WO 95/12619), which can be further oxidised to (partially) carboxylated units (see e.g. WO 00/26257). Nitroxyl-mediated oxidation using hypochlorite or a peracid results in 6-aldehyde- and 6-carboxy-anhydroglucose units (see e.g. WO 95/07303).

[0018] The oxidised glucans have improved water-solubility, altered viscosity and a retarded fermentability and can be used as metal-complexing agents, detergent additives, strengthening additives, bioactive carbohydrates, emulsifiers and water binding agents. They can also be used as starting materials for further derivatisation such as cross-linking and the introduction of hydrophobes. Oxidised glucans coupled to proteins can be used as emulsifiers and stabilisers. The oxidised glucans of the invention preferably contain 0.05-1.0 carboxyl groups, more preferably 0.2-0.8 carboxyl groups per anhydroglucose unit, e.g. as 6-carboxyl groups on 1,3-linked units.

[0019] When modified glucans with high proportion of carboxyl groups are desired, two oxidation processes can be combined or an oxidation can be combined with e.g. carboxymethylation (see below). Thus, an α -(1,3/1,6)-glucan having a degree of substitution (DS) for carboxyl groups between 0,3 and 1,0 can be conveniently prepared by first nitroxyl-mediated oxidation, resulting in 1,3-substituted units being oxidation to glucuronic acid units, followed by e.g. periodate and chlorite oxidation, resulting in 1,6-substituted units* being converted to ring-opened dicarboxy-substituted units. The order

of processes can also be inverted, or one oxidation process, such as nitroxyl-mediated 6-oxidation can be combined with carboxymethylation. Also, by appropriate adaptation of the oxidation processes mixed aldehyde-containing and carboxyl-containing polymers can be obtained.

- 5 [0020] Other useful modifications are alkylation, acylation, hydroxyalkylation, amino-alkylation, carboxyalkylation, phosphorylation, sulphatation, as well as physical and chemical crosslinking. Phosphorylation (see: O.B. Wurzburg (1986), Modified Starches: properties and uses. CRC Press Inc., Boca Raton, 97-112) can be achieved by dry heating glucans with a mixture of monosodium and disodium hydrogen phosphate or with tripoly-
10 phosphate. The phosphorylated glucans are suitable as wet-end additives in papermaking, as binders in paper coating compositions, as warp sizing-agents, and as core binders for sand molds for metal casting. Acylation, especially acetylation or propionylation using acetic or propionic anhydride respectively, results in products suitable as bleaching assistants and for the use in foils. Acylation with e.g. alkenyl succinic anhydrides or
15 (activated) fatty acids results in surface-active products suitable as e.g. surfactants, emulsifiers, and stabilisers. Crosslinking, e.g. by coupling oxidised derivatives, or by reaction with a crosslinking agent such as triphosphoric acid, epichlorohydrine or a dialdehyde, can be used to adjust the physical properties of the glucans, e.g. to enhance their water-binding or thickening capacities.
- 20 [0021] Hydroxyalkylation is commonly performed by base-catalysed reaction with alkylene oxides, such as ethylene oxide, propylene oxide or epichlorohydrin; the hydroxy-alkylated products have improved solubility and viscosity characteristics. Carboxy-methylation is achieved by reaction of the glucans with monochloroacetic acid or its alkali metal salts and results in anionic polymers suitable for various purposes including
25 crystallisation inhibitors, and metal complexants. Amino-alkylation can be achieved by reaction of the glucans with alkylene-imines, halo-alkyl amines or amino-alkylene oxides, or by reaction of epichlorohydrine adducts of the glucans with suitable amines. These products can be used as cationic polymers in a variety of applications, especially as a wet-end additive in paper making to increase strength, for filler and fines retention, and to
30 improve the drainage rate of paper pulp. Other potential applications include textile sizing and wastewater purification. The above mentioned modifications can be used either separately or in combination depending on the desired product. Furthermore, the degree of chemical modification is variable and depends on the intended use. If necessary 100% modification, i.e. modification of all anhydroglucose units can be performed. However,

partial modification, e.g. from less than 1 (e.g. 0.2) modified anhydroglucose unit per 100 units up to higher levels, will often be sufficient in order to obtain the desired effect.

[0022] Another suitable type of derivatives is formed by hydrolysates of the present glucans. Hydrolysis can be performed in a controlled manner in a way known per se, using e.g. dilute acid or glucanolytic enzymes, especially α -1,3-glucanases or α -1,6 glucanases. Hydrolysis results in polysaccharides of reduced chain length (degree of polymerisation, DP, of more than 20) or oligosaccharides (DP of less than 20).

[0023] The invention also relates to gluco-oligosaccharides containing the characteristic structure of the glucan described above. These can be produced using an isolated glucansucrase according to the invention or a *Lactobacillus* strain, or a recombinant micro-organism containing (a part of) a glucosyltransferase according to the invention. Gluco-oligosaccharides thus produced can be used as prebiotics and probiotics. The production of the gluco-oligosaccharides is different from the glucan synthesis reaction. In addition to sucrose, the substrate of the glucansucrase, an acceptor molecule such as maltose or lactose can be used as an acceptor, to synthesise oligosaccharides. Consecutive attachment of glucose units in a manner determined by the particular glucansucrase results in α (1,3)- and/or α (1,6)-linked gluco-oligosaccharides, the chain length of which can be determined by selecting the appropriate reaction conditions. Longer reaction times, higher sucrose levels and lower acceptor levels will usually result in relatively long chains, e.g. having a degree of polymerisation (DP) of more than 10, up to several hundreds if desired, while shorter reaction times, lower sucrose levels and higher acceptor levels will result in relatively short chains, e.g. with a DP from about 3 up to 10 or higher. Another way of producing gluco-oligosaccharides is by hydrolysis of the glucan described above. This hydrolysis can be performed by known hydrolysis methods such as enzymatic hydrolysis with enzymes such as amylase, dextranase or pullulanase or by acid hydrolysis. The produced gluco-oligosaccharides contain at least one 1,6- or one 1,3-glucosidic link to be used as prebiotics.

[0024] The invention also relates to a probiotic or synbiotic composition containing a *Lactobacillus* strain capable of producing a glucan and/or gluco-oligosaccharide according to the invention. The strain may also produce another poorly digestible poly- or oligosaccharide, such as a fructan. The probiotic or synbiotic compositions of the invention may be directly ingested with or without a suitable vehicle or used as an additive in conjunction with foods. They can be incorporated into a variety of foods and beverages including, but not limited to, yoghurts, ice creams, cheeses, baked products

such as bread, biscuits and cakes, dairy and dairy substitute foods, confectionery products, edible oil compositions, spreads, breakfast cereals, juices and the like.

[0025] Furthermore, the invention pertains to a process of improving the microbial status in the mammalian colon comprising administering an effective amount of a *Lactobacillus* strain capable of producing a glucan and/or gluco-oligosaccharide according to the invention. Furthermore, a process of improving the microbial status of the mammalian colon comprising administering an effective amount of a glucan or gluco-oligosaccharide according to the invention is also a part of the present invention.

10 **Examples**

General

The various lactic acid bacterial strains were isolated from a variety of sources, including fermented foods, the gastrointestinal tract of various human or animal species, and silage.

15 **Example 1: Identification and nucleotide sequence of glucansucrase/glucosyltransferase genes from lactobacilli**

The glucansucrase genes were identified by amplification with PCR using degenerated primers (GTFrev, 5' ADRTC NCCRT ARTAN AVNYK NG 3' and GTFforw, 5'-GAYAAAYWSNA AYCCNRYNGT NC-3'; N = A, C, G or T, Y = T or C, K = G or T, W = A or T, S = C or G, R = A or G), based on conserved amino acid sequences of different published glucansucrase genes. An amplification product with the predicted size of about 660 bp was obtained and cloned in *Escherichia coli* Top 10 using pCR-XL-TOPO (Invitrogen). Sequence analysis confirmed that part of a *gtf* gene had been isolated. The 660 bp amplified was used to design primers for inversed PCR. For inverse PCR chromosomal DNA was digested with 10 different enzymes ligated, yielding circular DNA molecules. PCR with the diverging primers with the circular ligation products as template yielded amplicons of various sizes, those products were cloned into pCR-XL-TOPO (Invitrogen) and sequenced (GATC, Konstanz, Germany). If necessary additional inverse PCR reactions were carried out to obtain the complete gene(s). Both strands of the entire glucansucrase genes were sequenced twice.

30 **Example 2: Isolation and identification of α -(1,6) glucan and a glucansucrase from *Lactobacillus reuteri* strain 180**

L. reuteri strain 180 was deposited as LMG P-18389 at the BCCM/LMG Culture Collection at Gent, Belgium. The strain was grown in 18 litres of MRS-s medium (in g per kg): yeast extract (22), sodium acetate trihydrate (5), sodium citrate dihydrate (2.42), ammonium chloride (1.32), dipotassium hydrogen phosphate (2), magnesium sulphate heptahydrate (0.2), manganese sulphate heptahydrate (0.05), sorbitan mono-oleate (1), vitamins (in mg per kg: B1: 14.4, B2: 3.6, B3: 72, H 0.216), sucrose (100), tap water

(remainder), for 21 h at 37°C under anaerobic conditions (pH 5.5). See also: Van Geel-Schutten et al., Appl. Microbiol. Biotechnol. (1998) 50, 697-703. During growth, 13 g/l polysaccharide was produced. This polysaccharide was isolated as described in the reference cited above. The monosaccharide composition of the polysaccharide was determined by hydrolysis of the soluble part of the polysaccharide and high-performance anion-exchange chromatography. It was characterised as a glucan. This glucan was not formed when the strain was grown on glucose instead of sucrose. Methylation analysis (Van Geel-Schutten et al. 1999) revealed the presence of 17-24% $\alpha(1,3)$ -linked glucosyl units, 34-44% of $\alpha(1,6)$ -linked glucosyl units, 7-15% of $\alpha(1,3,6)$ -linked glucosyl units and 7-35% of terminal glucosyl units. The average molecular weight of the glucan was determined to be 3.6×10^7 Da and the Rg was 45 nm.

The average molecular weight of the polysaccharide was established using the SEC-MALLS system: 0.0522 g of the glucan was dissolved in 10 ml DMSO/water (90/10) and heated for 1 hour at 80°C, filtered through a 0.45 μ m filter and injected on the SEC-MALLS system and analysed using the following conditions:

Eluent:	DMSO/water (90/10) with 0.1 M NaNO ₃
Flow rate:	0.5 ml/min
Injection volume:	0.247 ml
Column:	PLgel Guard, mixed-A and mixed-D
Temperature:	90°C

Detection: MALLS (DAWN-DSP), 50°C, $A_2=0$, $dn/dc=0.074$, F2 cell, RI; SDS PAGE followed by PAS-staining (Van Geel-Schutten et al. 1999) revealed the presence of an extracellular sucrase with a molecular weight of about 190 kDa. Part of the gene encoding the sucrase enzyme was isolated using PCR techniques and sequenced. On the deduced amino acid sequence of the fragment, high homologies were found with other glucan-sucrases. This partial sequence information is given in SEQ ID No. 1 (DNA) and 2 (protein). Full sequence information is given in SEQ ID No's. 11 and 12.

The glucan produced by *L. reuteri* strain 180 has been tested for application on ship hulls for the prevention of corrosion (see Example 8).

Example 3: Isolation and identification of $\alpha(1,6/1,3)$ glucan and a glucansucrase from *Lactobacillus reuteri* strain ML1

L. reuteri strain ML1, deposited as LMG P-20347 at the BCCM/LMG Culture Collection at Gent, Belgium, was grown overnight under anaerobic conditions at 37°C on MRS supplemented with sucrose (see Example 2). The cells were removed by centrifugation and two volumes of ethanol were added to the supernatant. The precipitated polysaccharides were harvested by centrifugation and resuspended in 2-3 liters of demi water and precipitated again with two volumes of ethanol. The glucan produced by this strain (7 g) was characterised by methylation analysis and monosaccharide composition analysis as

described in Example 2. The polymer was found to consist of 48-53% of $\alpha(1-3)$ linked glucosyl units, 3-8% of $\alpha(1-6)$ linked glucosyl units, 12-20% of $\alpha(1-3-6)$ linked glucosyl units (branching units) and 20-30% of 1-linked (terminal) glucose units. The glucans were not produced during growth on glucose. The average molecular weight of the polysaccharide was established to be 7.6×10^6 Da using the SEC-MALLS system as described in example 2. These were the first examples of the production of mutan-like polymers by lactobacilli. The glucan produced by *L. reuteri* strain ML1 has been tested for application as anticorrosive agent and showed excellent utility for the prevention of corrosion e.g. on ship hulls.

SDS PAGE followed by PAS-staining (Van Geel-Schutten et al. 1999) revealed the presence of an extracellular sucrase with a molecular weight of about 190 kDa. It was found that this strain produces two glucansucrases. Sequence information for these sucrase is given in SEQ ID No's 13 and 14 (ML1) and 15 and 16 (ML4).

Example 4: Isolation and identification of $\alpha(1,6/1,3)$ glucan and a glucansucrase from *Lactobacillus* strain LB 33.

A new *Lactobacillus* strain was obtained and was deposited as LMG P-20349. The strain was identified by 16S rRNA to be most closely related to *Lactobacillus parabuchneri*. The strain grown overnight on MRS supplemented with sucrose under anaerobic conditions at 37°C (see Example 2). 420 gram of glucan was produced. The glucan produced by this strain is not produced during growth on glucose.

Methylation analysis (see Example 2) revealed that the polymer consists of equal amounts of 29-39% of $\alpha(1-3)$ linked glucosyl units, 30-40% of $\alpha(1-6)$ linked glucosyl units, 3-13% of $\alpha(1-3-6)$ linked glucosyl units (branching units) and 15-30% of 1-linked (terminal) glucose units.

The average molecular weight of the polysaccharide was established to be 2×10^5 Da, using the SEC-MALLS system as described in Example 2.

By PCR with degenerated primers part of a sucrase type of glucosyl-transferase could be isolated indicating that the glucan is produced by a sucrase. This confirms the result that the glucan is produced during growth on sucrose and not on glucose. Part of the sucrase encoding gene was sequenced. On the deduced amino acid level high homologies were found with alternan sucrase from *Leuconostoc mesenteroides*. This indicates that the enzyme responsible for the glucan synthesis in *L. brevis* is the first alternan sucrase found in other bacteria than *Leuconostoc*. This partial sequence information is given in SEQ ID No. 3 (DNA) and 4 (protein). Full sequence information is given in SEQ ID No's. 17 and 18, respectively.

The glucan produced by this strain has thickening properties.

Example 5: Isolation and identification of α -(1,6) glucan and a glucansucrase from *Leuconostoc* strain 86

A new strain was obtained from silage and was deposited as LMG P-20350. The strain was identified by 16S rRNA to be a new *Leuconostoc* strain, most closely related to *Leuconostoc citreum*. The strain grown overnight on MRS supplemented with sucrose under anaerobic conditions at 37°C (see Example 2). 416 gram of glucan was produced. Methylation analysis of the glucan obtained revealed that more than 90 % of the glucose units was linked through an α (1,6) bond, identifying the polysaccharide as a dextran. The molecular weight of the glucan (determined as described in Example 2) was $3-4 \times 10^7$ Da and the Rg was 40 nm. The glucan is not produced during growth on glucose.

By PCR with degenerated primers 3 different fragments with part of a sucrase type of glucosyl-transferase could be isolated indicating that the glucan is produced by a sucrase and that possibly 3 sucrases are present in this strain. This confirms the result that the glucan is produced during growth on sucrose and not on glucose. Part of the sucrase encoding gene was sequenced. On the deduced amino acid level high homologies were found with DSRC and DSRB (fragment 1), alternan sucrase (fragment 2) and DSRA (fragment 3) from *Leuconostoc mesenteroides*. The sequence information is given in SEQ ID No's 5-10. *Leuconostoc citreum*, to which this new strain is most closely related, is not reported to produce dextran. The glucan produced by strain 86 has thickening properties.

Example 6: Identification of α -(1,6/1,3) glucan and a glucansucrase from *Lactobacillus sake* KG 15

Strain KG 15 was obtained from silage and was deposited as LMG P-21583. It was identified by 16S rRNA as *L. sake*. The strain was grown and the polysaccharide was recovered as described in example 2. The molecular weight of the polysaccharide was determined to be $4,7 \times 10^7$ Da (SEC MALLS) and the Rg was 92 nm. Methylation analysis (GC) revealed that the glucan produced by this strain is a largely linear dextran containing 4 % terminal glucose units, 86% of α (1,6) linked glucosyl units, 2% of α (1,3) linked glucosyl units and 8% α (1,3,6) disubstituted glucose units (branching points). The glucansucrase of this strain was sequenced (see SEQ ID No. 19 and 20).

Example 7: Identification of α -(1,6/1,3) glucan and a glucansucrase from *Lactobacillus fermentum* KG 3

Strain KG 3 was obtained from silage and was deposited as LMG P-21584. It was identified by 16S rRNA as *L. fermentum*. The strain was grown and the polysaccharide was recovered as described in example 2. The molecular weight of the polysaccharide was determined to be $2,4 \times 10^7$ Da (SEC MALLS) and the Rg was 107-119 nm. Methylation analysis (GC) revealed that the glucan produced by this strain is a largely linear dextran containing 3% terminal glucose units, 84% of α (1,6) linked glucosyl units,

8% of α (1,3) linked glucosyl units and 5% α (1,3,6) disubstituted glucose units (branching points). The glucansucrase of this strain was sequenced (SEQ ID No's 21 and 22).

5 **Example 8: Anticorrosion properties of glucans**

Plain carbon steel sheets of 1 cm² embedded in an epoxy matrix were exposed to a slightly corrosive medium (150 ml of 0.1 M LiClO₄) with or without the addition of a bacterial polysaccharide (0.2 g) for several days. The sheets were then examined visually and electrochemically from time to time. The corrosion potential (E_{corr} in mV with reference to Ag/AgCl) and polarisation resistance (R_p in k Ω /cm²) are both a measure of the anti-corrosion effect. After an initial adaptation of 3-10 hours, these parameters attained a stable value. The experiments were carried with a heteropolysaccharide from *Lactobacillus sake*, and a homopolysaccharide of the invention (from LB 180 according to example 4), as well as without polysaccharide. The results are summarised in the table below. It follows that the anti-corrosion properties of the glucan of the invention are superior. It was found that the homopolysaccharide of ML 1 (example 3) has at least equal anticorrosion performance as the LB 180 polysaccharide.

Table: Corrosion experiments

organism	type of polysaccharide	aspect of treated sheet	E_{corr} (mV vs. Ag/AgCl)	R_p (k Ω /cm ²)
control	-	corrosion	-700	1.5
<i>Lb. sake</i>	heteropolysaccharide	localised corrosion	-600	4.5
<i>Lb. 180</i>	α -glucan	thin black layer	-200	70

Example 9: Modification of α -1,3/1,6-glucan by oxidation

One gram (6.15 mmol of anhydroglucose units) of the α -1,3/1,6-glucan produced by strain LB 33 (example 4) is resuspended in 100 ml water. Next, 2,2,6,6-tetramethylpiperidine-1-oxyl (TEMPO; 0.01 g, 0.065 mmol) and sodium bromide (100 mg, 1 mmol) are added and the suspension is cooled to 0°C. The reaction can also be performed without bromide. A solution of hypochlorite (3 ml, 15% solution, 6.3 mmol) of pH 10.0 (0°C) is added. The pH is kept constant by addition of 0.1M NaOH. After 1 hr, the solution is poured into 150 ml 96% ethanol, causing the product to precipitate. The white precipitate is centrifuged, resuspended in ethanol/water (70/30 v/v) and centrifuged again. Next, the precipitate is resuspended in 96% ethanol, centrifuged and dried. The uronic acid content is determined by means of the uronic acid assay according to Blumenkrantz and Abdoe-Hansen (*Anal. Biochem.* 54 (1973), 484). A calibration curve was generated using polygalacturonic acid (5, 10, 15 and 20 μ g). With this calibration curve the uronic

acid content in a sample of 20 μ g of the product is determined. The major part of 6-hydroxyl groups have been oxidised to carboxyl groups.

Example 10: Construction of plasmids for expression of the glucansucrase genes in *E. coli*.

- 5 Two primers were designed with appropriate restriction sites; the C-terminal primer contained in all cases a His-tag. The PCR products were first cloned in pCR-XL-TOPO. The PCR products were removed from pCR-XL-TOPO using the appropriate enzymes and ligated in the appropriate sites of an expression vector (e.g pET15b (Novagen)).
- 10 For the expression of part of the glucosyltransferase gene of LB 180 (for better expression, the N-terminal region encoding the N-terminal variable domain of the glucansucrase, was not cloned) in *E. coli*, a PCR reaction was performed using Forw180 (5'-GATGCATGAG **CTCCCATGGG** CATTAAACGGC CAACAATATT ATTATTGACC C-3') containing *SacI* (bold) and *NcoI* (underlined) sites, and Rev180 (5'-ATATCGATGG GCCCCGGATC CTATTAGTGA *TGGTGATGGT* GATGTTTTTG
- 15 GCGTTTAAA TCACCAGGTT TTAATGG-3'), containing *ApaI* (bold), *BamHI* (underlined) and a 6x His-tag (*italics*) as primers. The PCR product was cloned in pCR-XL-TOPO. The PCR product was removed from pCR-XL-TOPO using *NcoI/BamHI* and ligated in the corresponding sites of pET15b (Novagen). The resulting plasmid (pET15b180) containing part of the glucansucrase gene of 704 amino acids encoding a glucansucrase without the variable N-terminal domain was transformed to *E. coli* B121
- 20 DE3 star (Invitrogen).

Cells of *E. coli* harbouring the pET15b180 were harvested by centrifugation after 16 h of growth under aerobic conditions at 37 °C. The pellet was washed with 50 mM sodium acetate buffer pH 5.5 containing 1 mM CaCl₂ and 1% (v/v) Tween 80 and the suspension

25 was centrifuged again. Pelleted cells were resuspended in with 50 mM sodium acetate buffer pH 5.5 containing 1 mM CaCl₂ and 1% (v/v) Tween 80, and 7.2 mM β -mercaptoethanol. Cells were broken by sonication and cell debris and intact cells were removed by centrifugation for 15 minutes at 4 °C at 14,000 rpm (Eppendorf). The resulting cell free extract was used as enzyme source to produce high molecular weight glucans from

30 sucrose in 50 mM sodium acetate buffer pH 5.5 containing 1 mM CaCl₂ and 1% (v/v) Tween 80 and 10 g/l sucrose. After 16 hours of incubation, the glucans were isolated using ethanol precipitation. When cell free extracts of *E. coli* B121 DE3 star (Invitrogen) harbouring the plasmid pET15b (without insert) were used as enzyme source, no glucans were produced from sucrose.

35

Sequence information

SEQ ID No's 1 and 2 give the nucleotide and amino acid sequence, respectively, of a part of the glucansucrase from strain Lb180 as originally determined (Example 2). The partial

sequence shows 53% (199/223) sequence identity and 68% similarity with dextransucrase DSRB742 of *Leuconostoc mesenteroides* (*Lc. mes.*), with 2 gaps (between amino acids F172 and N173), and 52% identity with some other dextransucrases and alternansucrases of *Lc. mes.*

- 5 SEQ ID No's 3 and 4 give the nucleotide and amino acid sequence, respectively, of a part of the glucansucrase from strain Lb 33 as originally determined (Example 4). The partial sequence shows 63% (143/224) sequence identity and 75% similarity with dextransucrase DSRB742 of *Lc. mes.* with 1 gap.

- 10 SEQ ID No's 5 and 6 give the nucleotide and amino acid sequence, respectively, of a part of a glucansucrase (86-1) from strain Lc 86 (Example 5). The partial sequence shows 98% (219/223) sequence identity and 99% similarity with dextransucrase DSRB742 of *Lc. mes.*

- 15 SEQ ID No's 7 and 8 give the nucleotide and amino acid sequence, respectively, of a part of another glucansucrase (86-5) from strain Lc 86 (Example 5). The partial sequence shows 55% (123/223) sequence identity and 68% similarity with dextransucrase DSRB742 of *Lc. mes.*, with 2 gaps (between amino acids M128 and R129 and between D162 and H163), and 51-56% identity with some other dextransucrases and alternansucrases of *Lc. mes.*

- 20 SEQ ID No's 9 and 10 give the nucleotide and amino acid sequence, respectively, of another glucansucrase (86-8) from strain Lc 86 (Example 5). The partial sequence shows 61-68% sequence identity and 74-78% similarity with dextransucrases and alternansucrases (including dextransucrase DSRB742) of *Lc. mes.*

- 25 SEQ ID No's 11 and 12 give the nucleotide and amino acid sequence, respectively, of the glucansucrase of strain Lb180 (Example 2). The sequence shows 1322/1768 (74%) sequence identity and 1476/1768 (82%) similarity with 15/1768 gaps with glucansucrase from *Lb. reuteri* LB 121 as disclosed in WO 01/90372. The -35 and -10 sites TTGAAA and TATAA are located at nucleotide positions 561 and 599, respectively. The ribosome binding site (RBS) GAAGGAG is at 574 and the start codon ATG at 587. Inverted repeats AAGCAGCTC and GAGCTGCTT are at 6025 and 6051. Possible stop codons (TAA, TAG, TGA) are indicated with an * (5963).

- 30 SEQ ID No's 13 and 14 give the nucleotide and amino acid sequence, respectively, of the glucansucrase I from strain ML1 (Example 3). The sequence shows 1327/1775 (74%) sequence identity and 1465/1775 (81%) similarity with 17/1775 gaps with glucansucrase from *Lb. reuteri* LB 121 as disclosed in WO 01/90372, and 43-44% sequence identity and 57-58% similarity with dextransucrases of *Lc. mes.* and 47% sequence identity and 61% similarity with an alternansucrases of *Lc. mes.* The RBS AAGGAGA is at 31 and the start codon ATG is at 43. A stop codon TAG is at 5356.

35 SEQ ID No's 15 and 16 give the partial nucleotide and amino acid sequence, respectively, of a second glucansucrase from strain ML1 (ML4) (Example 3). The sequence shows

301/817 (36%) sequence identity and 427/817 (51%) similarity with 12/817 gaps with glucansucrase from *Lb. reuteri* LB 121 as disclosed in WO 01/90372, and 38% sequence identity and 53% similarity with glucosyltransferase of *Streptococcus mutans*.

5 SEQ ID No's 17 and 18 give the partial nucleotide and amino acid sequence, respectively, of the glucansucrase from strain LB 33 (Example 4). The sequence shows 59% sequence identity and 71% similarity with several known dextransucrases of *Lc. mes.* and 53% sequence identity and 67% similarity with other known dextransucrases (including dextransucrase DSRB742) of *Lc. mes.*

10 SEQ ID No's 19 and 20 give the nucleotide and amino acid sequence, respectively, of the glucansucrase from *Lb.* strain KG 15 (Example 6). The sequence shows 496/1111 (44%) sequence identity and 637/1111 (56%) similarity with 71/1111 gaps with glucansucrase from *Lb. reuteri* LB 121 as disclosed in WO 01/90372, and 57-59% sequence identity and 70% similarity with several dextransucrases (including dextransucrase DSRB742) of *Lc. mes.* The -35 and -10 sites *TTGGAC* and *TATTAT* are located at nucleotide positions 477 and 502, respectively. The RBS *GAAAGGA* is at 593 and the start codon *ATG* at 608. A stop codon *TAG* is 5393. Inverted repeats *AAAACAACCCCC* and *GGGGTTGTTTTT* are at 5497 and 5531 (-10.7 kcal/mole).

15 SEQ ID No's 21 and 22 give the partial nucleotide and amino acid sequence, respectively, of the glucansucrase from *Lb.* strain KG 3 (Example 7). The sequence shows 58 sequence identity and 71% similarity with known dextransucrases (including dextransucrase DSRB742) of *Lc. mes.*

Description of the figure

25 Figure 1 depicts an amino acid sequence alignment of glucosyltransferases (GTF) according to the invention. It shows the partial sequences of the GTF of Lb 180 (first line, starting with amino acid 216 of SEQ ID No. 12); GTF of ML1 (second line, starting with amino acid 15 of SEQ ID No. 14), GTF of Lb 33 (third line, starting with amino acid 222 or 243 of SEQ ID No. 18); GTF of KG15 (fourth line, starting with amino acid 567 of SEQ ID No. 20) and GTF of KG3 (fifth line, starting with amino acid 1 (LMAAF) of SEQ ID No. 22); and a GTF according to the invention of a *Lb. reuteri* strain "104" (sixth line, 1 (WPNTV) - 525). The alignment is not necessarily the best fit according to automated alignment programs, but is intended to define the enzymes of the invention.

35 The invention not only covers amino acid sequences shown in this figure, but also sequences wherein amino acids of a given sequence in the figure are exchanged with the corresponding amino acids (including gaps) of another sequence of the figure. This applies to stretches of at least 100 amino acids having at least 80%, preferably at least 90% identity with any of the sequences of the figure, or of the sequences listings given separately. It especially applies to the stretch of amino acids between the consensus peptides DNSN and YYGD (from 1202 to 1422 of SEQ ID No 12). Especially preferred

are sequences comprising the active core of the enzymes, which are present between the consensus peptides INGQ and VPDQ (from 957 to 1724 of SEQ ID No 12), with preferably at least 70% identity with any one of the core sequences given. A preferred non-identity with a given sequence is an exchange with the corresponding amino acids of another sequence. Especially preferred sequences are those where an amino acid at a given position is shared between at least 2, in particular at least 3, of the sequences of the figure. Most preferred are those sequences in which one of those consensus sequences is that of the GTF of Lb180, ML1 or Lb33 (first three lines). The N-terminal part upstream of the core (shown in the figure for GTF 180 and GTF ML1 only), or the C-terminal part downstream of the core (not shown in the figure) may be wholly or partly present or may be absent.

Claims

1. A process of producing a glucan having at least 10 anhydroglucose units, having a backbone consisting essentially of $\alpha(1,3)$ - and/or $\alpha(1,6)$ -linked anhydroglucose units (AGU), comprising subjecting sucrose to the activity of a glucosyltransferase produced by a *Lactobacillus* strain capable of producing $\alpha(1,3)$ - and/or $\alpha(1,6)$ -linked glucans, or to the *Lactobacillus* strain capable of expressing the glucosyltransferase.
2. A *Lactobacillus* strain capable of producing, in the presence of sucrose, a glucan having at least 10 anhydroglucose units (AGU) having a backbone consisting essentially of $\alpha(1,3)$ - and/or $\alpha(1,6)$ -linked AGU.
3. A glucan capable of being produced by glucosyltransferase activity of a lactic acid bacterium on a sucrose substrate, the glucan having an average molecular weight between 10 kDa and 1 GDa, especially between 10kDa and 50 MDa, and having a backbone consisting essentially of $\alpha(1,3)$ - and $\alpha(1,6)$ -linked anhydroglucose units (AGU).
4. A glucan according to claim 3, which is capable of being produced by glucosyltransferase activity of a *Lactobacillus* species.
5. A glucan according to claim 4, comprising 15-80% of $\alpha(1,3)$ -linked AGU, 2-80% of $\alpha(1,6)$ -linked AGU, and 2-25% of $\alpha(1,3,6)$ -linked AGU.
6. A glucan according to claim 5, having an average molecular weight of 50 kDa - 1 MDa and comprising 30-45% of $\alpha(1,3)$ -linked AGU, 30-45% of $\alpha(1,6)$ -linked AGU, and 3-13% of $\alpha(1,3,6)$ -linked AGU.
7. A glucan according to claim 5, having an average molecular weight of 10-50 MDa and comprising 15-26% $\alpha(1,3)$ -linked AGU, 30-50% of $\alpha(1,6)$ -linked AGU, 5-20% of $\alpha(1,3,6)$ -linked AGU.
8. A glucan according to claim 5, having an average molecular weight of 1-50 MDa and comprising 45-60% of $\alpha(1,3)$ -linked AGU, 4-10% of $\alpha(1,6)$ -linked AGU, and 10-20% of $\alpha(1,3,6)$ -linked AGU.

9. A glucan capable of being produced by glucosyltransferase activity of a lactic acid bacterium on a sucrose substrate, having an average molecular weight of 10-50 MDa and comprising 80-99% of $\alpha(1,6)$ -linked AGU and 0-15% of $\alpha(1,3)$ -linked AGU.
10. A protein having glucosyltransferase activity, capable of producing, in the presence of sucrose, a glucan according to any one of claims 3-9.
11. A protein according to claim 10, comprising an amino acid sequence of at least 100 amino acids exhibiting at least 70%, preferably at least 80%, amino acid identity with any one of the amino acid sequences of SEQ ID No. 2, 4, 8, 10, 12, 14, 16, 18, 20 and 22, and/or having a stretch of 100 amino acids having at least 80%, preferably at least 90%, amino acid identity with any one of the said amino acid sequences, or having at least 99% amino acid identity with the amino acid sequence of SEQ ID No. 6, and/or having a stretch of 100 amino acids having 100% amino acid identity with the amino acid sequence of SEQ ID No. 6.
12. A nucleic acid sequence encoding a protein according to claim 11.
13. A recombinant host cell containing one or more copies of a nucleic acid construct comprising a nucleic acid sequence according to claim 12 and capable of expressing a protein having glucosyl-transferase activity.
14. A *Lactobacillus* strain, capable of producing a glucan according to any one of claims 3-9, especially a *Lactobacillus* strain corresponding to strain 33, 180 or ML1 as described herein.
15. A *Leuconostoc* strain, capable of producing a glucan according to claim 9, especially a *Leuconostoc* strain corresponding to strain 86, deposited under accession number LMG P-20350.
16. A chemically modified glucan, which is obtained by 2,3-oxidation, 6-oxidation, phosphorylation, acylation, alkylation, hydroxyalkylation, carboxymethylation, amino-alkylation of one or more AGU of a glucan according to any one of claims 3-9.
17. Use of a glucan according to any one of claims 3-9, as a thickener.
18. Use of a glucan according to any one of claims 3-9, as a prebiotic and/or as a bioactive agent.

19. Use of a glucan according to any one of claims 3-9, as an anti-corrosion agent.
20. Use of a *Lactobacillus* bacterium capable of producing a glucan according to any one of claims 3-9, as a probiotic agent, or together with an indigestible glucan, as a synbiotic agent.

FIG. 1 SEQUENCE ALIGNMENT

216 MEIKKHFKLYKSGKQWVTA AVATVAVSTALLYGGVAHADQQVQSSTTQEQTSTVNADTTK
 15 MEIKKHFKLYKSGKQWVTA AVATVAVSTALLYGGVAHADQQVQSSTTQDQTSTVNTNTTK

 276 TVNLDTNTDQPAQTDDKNQVANDTTTNQSKTDSTSTTVKNPTFIPVSTLSSSDNEKQSQN
 75 TIAADTNADQPAQTADKNQAASNDTTTNQSKTDSTSTTVKNLTSTFPVSTLPSTDNEKQSQN

 336 YNKPDNGNYGNVDAAYFNNNQLHISGWHATNASQGTDSRQVIVRDITTKTELGRNTVNTNN
 135 YNKHDNGNYGNIDTAYFSNNQLHVS GWNATNASQGTNSRQIIVRDITTNNELGRTDVTNN

 396 VLRPDVKNVHNVYNADNSGFDVNINIDFSKMKDYRDSIEIVSRYSNGNGKSVDWWSQPITF
 195 VARPDVKNVHNVYNADNSGFDINVNIEFSKMKDYRDSIEIVSRYSNGNGKSIDWWSQPITF

 456 DKNNYAYLDTFEVKNGELHATGWNATNKAINYNHHFVILFDRTNGKEVTRQEV RDGQSRP
 255 DKNNYAYLDTFEVKNGELHATGWNATNSAINYNHHFVILFDQTNGKEVARQEVREGQSRP

 516 DVAKVYPQVVGANN SGFDVTFNIGDL DYTHQYQILSRYSNADNGEGDYV TYWFAPQSIAP
 315 DVAKVYPQVVGADNSGFDVTFNIGNLDYTHQYQVLSRYSNSDNGEGDNV TYWFPQSIAP

 576 ANQSNQGYLDSFDISKNGEVTVTGWNATDLSELQTNHYVILFDQTAGQQVASAKVDLISR
 375 ANQSNQGYLDSFDISKNGEVTVTGWNATDLSELQNNHYVILFDQTAGKQVASAKADLISR

 636 PDVAKAYPTVKTAETSGFKVTFKVSNLQPGHQYSVVS RFSADENGNGNDRHTDYWYSPV
 435 PDVAKAYPTVKTAANS GFKVTFKVN DLQPGHQYSVVS RFSADENGNGNDRHTDYWYSPV

 696 TLNQ TASNIDTITMTS NGLHITGWMASDNSINEATPYAIILNNGREVTRQKLT LIARPDV
 495 TLNQ NASNIDTITMTS NGLHIGSWMASDNSINETTPYAIILNNGKEVTRQKMSLTARPDV

 756 AAVYPSLYNSAVSGFDTTIKLTNAQYQALNGQLQVLLRFSKAVDGNPNGTNTVTDQFSKN
 555 AAVYPSLYNSAVSGFDTTIKLTNDQYQALNGQLQVLLRFSKAADGNPSGDNTVTDQFSKN

 816 YATTGGNFYDVKVNGNQIEFSGWHATNQSN DKNSQWIIVLVNGKEVKRQLVNDTKDGAAG
 615 YATTGGNFYDVKVNGNQVEFSGWHATNQSN DKDSQWIIVLVNGKEVKRQLVNDTKEGAAG

 876 FNRNDVYKVNPAIENSIMSGFQGIITLPVTVKDEN VQLVHRFSNDAKTGEGNYVDFWSEV
 675 FNRNDVYKVNPAIENS SMSGFQGIITLPVTVKNE NVQIVHRFSNDAKTGEGSHVDFWSEV

 936 MSVKDSFQKGNGLNQFGLQTINGQYYIDPTTGQPRKNFLLQNGNDWIYFDKDTGAGTN
 735 MPVKDSFQKGNGLKQFGLQTINGHQQYYIDPMTGQPRKNFLLQNGNDWLYFDNETGEGTN
 222 VNGKIYFVGDNQVKKNF TAIINGQSLYFNKTTGELASNDVQYENGLVKINDV
 567 QTIAGKTY YFDKD GHLRKGYSTIIDNQLYYFDLKTGESVS

 996 ALKLQFDKGTISADEQYRRGNEAYS YDDKSIENVNGYLTADTWYRPKQILKDGT TWTDSK
 795 ALKRQFDGGTISADSQYRK GNEAYGDNKSIENVDGFLTADTWYRPKQILKW TWTDSK
 275 HNAAYSIDP?GFTNVNGFLTANSWYRPKYIYKDGQKQWVEST
 607 TTTSNFKSGLTSQTDDTTPHNSAVNMSKDSFTTVDGFLTAE SWYVPKDIQTSATDWRAS T

 1056 ETDMPILMVWWPNTVTQAYYLN YMKQYGNLLPASLP SFSTADSAELNHYSELVQQNIE
 854 ETDMPRLLMVWWPNTVTQAYYLN YMKQHGNLLPANLPFFNSDADPLELNYYAEIVQQNIE
 316 SQDMPRLLMTW PDKNTQVAYLQYM QKMGI LPADVTISSQTNQSVLTKE SFITQAEIE
 666 PEDFRPIMMTW WP TKQIQAYYLNH MVSEG LLSSDKKFSATD DQTL LNQA AHAVQLQIE
 (0)
 1 WPNTVTQAYYLN YMKQHGNLLPASLPFFNADADPAELNHYSEIVQQNIE

1116 KRISSET GSTDWLRTLMEHFVTKNSMWNKDSENVYGGGLQLQGGFLKYVNSDLTKYANSW
914 KKISQT GNTDWLRTLMEHFVSNNTMWNKSENEDEFGGLQLQGGFLKYVNSDKTPNANSNW
374 KQIGVTNGNTDWLKKDISDFVNSQPNWNIDSEAKGTDH LQGGALLYVNKLTYPYANSY
725 LKIQQT KSVEWLRTTMHNFIKSQPGYNVTSETPSNDH LQGGALSYINSVLTDPDANSNF
1 LMAAFVVTQPQWNKTSEDVNDDH LQGGALT FENNGDT DANDSY
50 KRISSET GNTDWLRTLMEHFVTKNSMWNKDSENVYGGGLQLQGGFLKYVNSDLTKYANSW

1176 RLMNRTATNIDGKNY GGAEFLLANDIDNSNPVVQAEELNWLYYLMNFGTITGN
974 RIMGRQPANIDGNP IGSEFLLANDVDNSNPVVQAEQLNWLHYLLNFGTITAN
433 RLLNRTLNTQQGQVKDTS KQGGYEMLLANDVDNSNPVVQAEQLNWLYYMMNIGSITAN
783 RLMNRRNPTQQDGTRHYNTDTSSEGGYELLLANDVDNSNPVVQAEQLNWLHYFLTHFGEIVKN
44 RLMNRTPTNQTGERLYHIDDSLGGYELLLANDVDNSNPQVQAEQLNWLHYLMHFGDITAD
110 RLMDRATATNIDGKNY GGAEFLLANDIDNSNPVVQAEELNWLYYLMNFGTITGN

1229 NPEANFDGIRVDAVDNVDVLLSIARDYFNAAYNMEQSDASANKHINILEDWGWD DPAYV
1027 DPDANFDSIRVDAVDNVDADLLDIAGDYFNAVYHSQSNDKIANAHINILEDWGQDPYYT
491 DPTANFDGYRVDVDNVDADLLNIAADYAKAYKTN QSDANANKHLSILEDWDNDNDPAYI
843 DPSANFDSVRVDVDNVDADLLNITAAFRDVGVDKNDLTANQHLSILEDWGHNDDPLYV
104 DPDANFDAIRIDVDNVDADLLQLAAQYFRDAYGMATTDATSNKHLSILEDWSHNDPAYM
163 NPEANFDGIRVDAVDNVDVLLSIARDYFNAAYNMEQSDANANKHINILEDWGWD DPAYV

1289 NKIGNPQLTMDDRLRNAIMDTLSGAPDKNQALNKLITQSLVNRANDN TENAVIPSYNFV
1087 QSIGTPQLSMDYNFSTIRSVLASNTASMTD IIKNSLVNRSLDN AENVSI PNYSFI
551 KAHGNNQLTMDFPAHLAIKYSLNMPVSQSRGLEPELTTSLVNRTGDDSTENVAQPNYTFI
903 KDHGSDQLTMDDYMHTQLIWSLTKNPDNRSAMRRFMEYYLVDRADN TSDPAIPNYSEFV
164 QAHGNDQLTMDDYMHTQLIWSLTKEAQRGTMARFMDFYLTNRANDD TENTAQPSYSFV
223 NKIGNPQLTMDDRLRNAIMDTLSGAPDKNQALNKLITQSLVNRANDN TENAVIPSYNFV

1348 RAHDSNAQDQIRQAIQAATGKPYGE FNLDDEKKGMEAYINDQNSTNKKWNLYNMPSAY
1142 RAHDNGSQDDIKRAISDVNNLPYGSK FNFEQEQKGIEAYIADQSNVNKKWNNYNI PSSY
611 RAHDSEVQTIIAQIIKDKINPNSDGLTVPDEISQAFKIYNADELKTDKQYTFYNMPSAY
962 RAHDSEVQTVIGDIVAKLYPDVKNSL PSMEQLAAAFKVYDADMNSVNKKYTQYNMPAAY
223 RAHDSEVQTVIAEIVTKLHPEAGNGLMPTEEQMAEAFKIYNADQKKAVKTYTHYNMPSAY
282 RAHDSNAQDQIRQAIQAATGKPYGE FNLDDEKKGMEAYINDQNSTNKKWNLYNMPSAY

1406 TILLTNKDSVPRVYGGDLYQDGGQYMEHKTRYFDTITNLLKTRVKYVAGGQTM SVDKN
1201 AIMLTNKDTPRVYGGDLFTDGGQYMAQTTRYYPALTSLLKARIKYVAGGQTM SVDKN
671 TILLTNKDTPRVYGGDLYSDNGNYMSAHPYDAITTLKTRMKYVSGGQNM RMQYMQG
1021 AMLLTNKDTPRVYGGDMYTDGQYMATKSPYDAISALLKARIKYVAGGQTM MAVDKH
283 AMLLTNKDVIPRIYGGDLYTDGQYMATKSPYFDAISTMLQARTKYVAGGQTM MAVDQH
340 TILLTNKDSVPRVYGGDLYQDGGQYMEHKTRYFDTITNLLKTRVKYVAGGQTM SVDKN

1464 GILTNVRFKGKAMNATDTGTDETRTEGIGVVISNNTNLKLN DGESVVLHMG
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 Phe Asp Gly Tyr Arg Val Asp Ala Val Asp Asn Val Asp Ala Asp Leu
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 65 70 75 80
 Asn Asn Asp Pro Ala Tyr Ile Lys Ala His Gly Asn Asn Gln Leu Thr
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 His Asn Asp Pro Glu Tyr Val Lys Asp Phe Gly Asn Asn Gln Leu Thr
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 Asn Arg Asn His Asp Ser Thr Glu Asn Thr Ala Ile Pro Asn Tyr Ser
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 Phe Val Arg Ala His Asp Ser Glu Val Gln Thr Val Ile Ala Gln Ile
 145 150 155 160
 Ile Ser Glu Leu His Pro Asp Val Lys Asn Ser Leu Ala Pro Thr Ala
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 Asp Gln Leu Ala Glu Ala Phe Lys Val Tyr Asn Asn Asp Glu Lys Gln
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Ser Asp Val Asn Ala Asn Lys His Ile Ser Ile Leu Glu Asp Trp Ser
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Gly Leu Asp Pro Asn Glu Val Val Lys Asn Gly Asn Pro Gln Leu Thr
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Phe Ile Arg Ala His Asp Ser Glu Val Gln Thr Val Ile Ala Gln Ile
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Ile Lys Asp Lys Ile Asn Thr Lys Ser Asp Gly Leu Thr Val Thr Pro
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SEQ ID No. 12 PRT

Lactobacillus reuteri strain 180

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101 I L S Q C S I I A N F L L G K K V Y C G

361 AAATATTTAAGAATATTGTCGTTACCGGTAGAGACAATTTTATAAGTTCTAACTTTGTTC
121 N I * E Y C R Y R * R Q F Y K F * L C S

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141 L C C * P L L G S * T Y Y G F R * V N L

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- 35

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341 N G N Y G N V D A A Y F N N N Q L H I S

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1381 CGCATACCTTGACACATTTGAAGTTAAAAATGGGGAATTGCATGCAACAGGATGGAATGC
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2221 AGAGGTTACTCGTCAAAAATTAACCTTAATTGCGCGTCCAGATGTAGCAGCAGTATATCC
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2761 TGCAAAGACTGGTGAAGGTAATTATGTTGATTTCTGGTCAGAAGTAATGTCTGTTAAGGA
921 A K T G E G N Y V D F W S E V M S V K D

2821 CAGCTTCCAAAAGGGTAATGGTCCGCTTAATCAATTTGGTTTACAACTATTAACGGCCA
941 S F Q K G N G P L N Q F G L Q T I N G Q

2881 ACAATATTATATTGACCCAACAACCTGGCCAACCTCGTAAGAATTTCTTTATTGCAAAATGG
961 Q Y Y I D P T T G Q P R K N F L L Q N G

2941 GAACGATTGGATTTACTTTGACAAAGATACTGGTGCTGGAACATAATGCTCTTAAGTTACA
981 N D W I Y F D K D T G A G T N A L K L Q

3001 ATTTGATAAGGGAACAATTTCTGCTGATGAGCAATATCGTCGAGGAAATGAAGCCTATAG
1001 F D K G T I S A D E Q Y R R G N E A Y S

3061 TTATGATGACAAGAGTATTGAAAATGTAAATGGTTACTTAAACAGCTGATACTTGGTACCG
1021 Y D D K S I E N V N G Y L T A D T W Y R

3121 ACCAAAACAAATCTTAAAGGATGGTACTACTTGGACTGACTCTAAAGAAACAGATATGCG
1041 P K Q I L K D G T T W T D S K E T D M R

3181 CCCAATTTTAATGGTATGGTGGCCAAATACTGTTACACAAGCATATTATCTTAACCTACAT
1061 P I L M V W W P N T V T Q A Y Y L N Y M

3241 GAAGCAATATGGTAATTTATTGCCGGCTAGTTTACCAAGCTTCAGTACAGATGCTGATTC
1081 K Q Y G N L L P A S L P S F S T D A D S

3301 TGCTGAATTAAATCATTACTCCGAGCTTGTTCAACAAAATATCGAAAAGCGGATCAGTGA
1101 A E L N H Y S E L V Q Q N I E K R I S E

3361 GACTGGTAGTACTGATTGGTTACGTACACTAATGCATGAGTTCGTTACTAAGAATTCTAT
1121 T G S T D W L R T L M H E F V T K N S M

3421 GTGGAATAAGGATAGTGAAAATGTCGATTACGGTGGTTTGCAATTACAAGGTGGATTCCCT
1141 W N K D S E N V D Y G G L Q L Q G G F L

3481 TAAGTATGTAAATAGTGATCTTACTAAATATGCAATTCAGATTGGCGTTTAAATGAACCG
1161 K Y V N S D L T K Y A N S D W R L M N R

3541 TACAGCTACTAATATTGATGGTAAGAACTATGGTGGTGCGGAATTCTTATTAGCTAATGA
1181 T A T N I D G K N Y G G A E F L L A N D

3601 TATTGATAACTCAAATCCAGTTGTTCAAGCTGAAGAATTAACTGGCTTTACTATTTAAT
1201 I D N S N P V V Q A E E L N W L Y Y L M

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3661 GAATTTTCGGTACAATTACAGGAAATAATCCTGAAGCTAATTTTGATGGTATTCGAGTGGGA
1221 N F G T I T G N N P E A N F D G I R V D

3721 TGCTGTGATAATGTAGATGTTGACTTATTGAGTATTGCACGTGATTACTTTAATGCAGC
1241 A V D N V D V D L L S I A R D Y F N A A

3781 ATATAACATGGAGCAAAGTGATGCCAGTGCTAATAAGCACATTAATATTTTGGGAAGATTG
1261 Y N M E Q S D A S A N K H I N I L E D W

3841 GGGATGGGATGATCCTGCTTATGTAAATAAGATTGGAAATCCTCAATTAACAATGGATGA
1281 G W D D P A Y V N K I G N P Q L T M D D

3901 TCGTTTACGAAATGCAATTATGGATACATTATCAGGAGCACCTGATAAAAACCAAGCATT
1301 R L R N A I M D T L S G A P D K N Q A L

3961 GAATAAATTAATTACTCAGTCATTAGTAAATCGTGCTAATGATAATACTGAAAACGCGGT
1321 N K L I T Q S L V N R A N D N T E N A V

4021 TATTCCAAGCTATAATTTTGTTCGAGCACATGATAGTAATGCTCAAGACCAAATTCGTCA
1341 I P S Y N F V R A H D S N A Q D Q I R Q

4081 GGCTATTCAAGCTGCAACTGGAAAACCATATGGCGAATTTAACTTAGATGATGAAAAGAA
1361 A I Q A A T G K P Y G E F N L D D E K K

4141 GGGTATGGAAGCATATATTAATGATCAGAATCTACTAATAAGAAGTGAATCTTTACAA
1381 G M E A Y I N D Q N S T N K K W N L Y N

4201 TATGCCTTCTGCTTATACTATTCTTCTAACAATAAAGATTCAGTTCCTCGTGTTTACTA
1401 M P S A Y T I L L T N K D S V P R V Y Y

4261 TGGAGACCTCTACCAAGATGGTGGTCAATATATGGAACATAAAACACGTTACTTTGATAC
1421 G D L Y Q D G G Q Y M E H K T R Y F D T

4321 TATTACTAACTTATTAAAGACACGGGTAAATATGTTGCCGGTGGACAAACTATGAGTGT
1441 I T N L L K T R V K Y V A G G Q T M S V

4381 TGATAAGAATGGTATTCTTACAAACGTTCTGTTTTGGGAAAGGCGCCATGAATGCTACTGA
1461 D K N G I L T N V R F G K G A M N A T D

4441 TACTGGTACTGATGAAACAAGAACAGAAGGTATCGGTGTTGTAATTAGTAACAATACTAA
1481 T G T D E T R T E G I G V V I S N N T N

4501 TTTGAAGCTTAATGATGGTGAATCAGTAGTGCTTCATATGGGAGCTGCTCATAAGAATCA
1501 L K L N D G E S V V L H M G A A H K N Q

4561 AAAGTATCGTGCTGTGATCTTAACAACCTGAAGATGGTGTTAAGAATTACACTAATGATAC
1521 K Y R A V I L T T E D G V K N Y T N D T

4621 AGACGCACCAAGTTGCATACACTGATGCTAATGGTGACCTTCACTTTACTAATACTAATTT
1541 D A P V A Y T D A N G D L H F T N T N L

4681 AGATGGTCAACAATATACAGCTGTTTCGTGGATATGCAAATCCTGATGTAACAGGATATCT
1561 D G Q Q Y T A V R G Y A N P D V T G Y L

4741 AGCTGTTTGGGTACCAGCTGGAGCAGCAGATGATCAAGATGCACGTACTGCACCAAGTGA
1581 A V W V P A G A A D D Q D A R T A P S D

4801 TGAGGCCCATACTACAAAGACTGCTTATCGCTCTAATGCAGCCCTTGATTCTAACGTTAT
1601 E A H T T K T A Y R S N A A L D S N V I

4861 TTATGAAGGATTCTCTAACTTCATTTACTGGCCAACCTACTGAAAGCGAACGGACTAATGT
1621 Y E G F S N F I Y W P T T E S E R T N V

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4921 GAGAATTGCACAAAATGCGGATCTATTTAAGTCATGGGGAATTACTACCTTTGAATTAGC
1641 R I A Q N A D L F K S W G I T T F E L A

4981 TCCACAATACAATTCAAGTAAAGATGGTACGTTCCCTTGATTCAATAATTGATAATGGATA
1661 P Q Y N S S K D G T F L D S I I D N G Y

5041 TGCCTTTACTGATCGTTATGATTTAGGAATGAGTACTCCTAACAAGTATGGATCTGATGA
1681 A F T D R Y D L G M S T P N K Y G S D E

5101 AGACTTACGTAATGCTTTACAAGCCTTACATAAAGCTGGTTTACAAGCAATTGCCGACTG
1701 D L R N A L Q A L H K A G L Q A I A D W

5161 GGTTCCTGATCAAATTTATAACTTACCTGGTAAAGAAGCTGTAACAGTAACACGTTTCAGA
1721 V P D Q I Y N L P G K E A V T V T R S D

5221 TGATCACGGTACTACATGGGAAGTTTCGCCAATAAAGAATGTTGTCTATATTACAAATAC
1741 D H G T T W E V S P I K N V V Y I T N T

5281 GATTGGTGGAGGTGAATACCAGAAGAAATATGGTGGTGAATTCTTAGACACTCTTCAAAA
1761 I G G G E Y Q K K Y G G E F L D T L Q K

5341 AGAATATCCACAATTATTTAGTCAGGTATATCCAGTAACTCAAACGACAATTGATCCTAG
1781 E Y P Q L F S Q V Y P V T Q T T I D P S

5401 TGTTAAGATTAAAGAGTGGTCTGCTAAATACTTTAATGGTACTAATATCCTTCATCGAGG
1801 V K I K E W S A K Y F N G T N I L H R G

5461 TGCTGGATATGTATTGCGCTCTAATGATGGTAAATACTATAATCTTGGTACAAGCACTCA
1821 A G Y V L R S N D G K Y Y N L G T S T Q

5521 ACAATTCTTACCGTCTCAATTATCAGTTCAAGATAATGAAGGATATGGATTTGTAAAAGA
1841 Q F L P S Q L S V Q D N E G Y G F V K E

5581 AGGAAATAATTACCATTACTATGATGAGAATAAACAGATGGTAAAAGATGCGTTTATTCA
1861 G N N Y H Y Y D E N K Q M V K D A F I Q

5641 AGATAGTGTGTTGGTAATTGGTATTACTTCGATAAAAATGGTAATATGGTTGCTAACCAAAG
1881 D S V G N W Y Y F D K N G N M V A N Q S

5701 TCCTGTTGAAATTAGTAGTAATGGAGCTTCAGGAACTTACCTTTTCTTGAACAATGGGAC
1901 P V E I S S N G A S G T Y L F L N N G T

5761 ATCATTCGTTCTGGATTGGTGAAACTGATGCAGGTACGTACTATTATGATGGCGATGG
1921 S F R S G L V K T D A G T Y Y Y D G D G

5821 CCGAATGGTTCGTAATCAAACGGTAAGTGATGGTGCGATGACATATGTTCTTGATGAAAA
1941 R M V R N Q T V S D G A M T Y V L D E N

5881 TGGTAAACTTGTTAGTGAATCATTTGATTCTGCTACTGAAGCACACCCATTAAAACC
1961 G K L V S E S F D S S A T E A H P L K P

5941 TGGTGATTTAAACGGCCAAAAATAATTACAATATGAAAATTGGAACCTTGATTTTACCTT
1981 G D L N G Q K * L Q Y E N W N L Y F T F
inverted repeat

6001 CTTTGAAATAATATAGTTCTAATTAAGCAGCTCGCACCAAGACTTGGTATGAGCTGCTTT
2001 F E I I * F * L S S S H Q D L V * A A F

6061 TTTTGGCTCTACAATATCTGGTGTGATATAGAAATATCACTTTCTATACCAATATCAGA
2021 F G S T I S G V D I E I S L S I P I S D

6121 TTTTGTTTTTTAAACTAAAAAGAGGCTCGCCCTCTGATACAATGAAATCGCCAAATCAC
2041 F C F * T K K E A R P L I Q * N R Q I T

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6181 ATAGTAAAGAAGGTAACCTCCATGGATAATGATACAAGAAGCTCTTCTCAATTTAACAGAC
2061 * * R R * P P W I M I Q E L F S I * Q T

6241 CCTCATTTAAATTTTCCTCATCATTGGCTTAAATATAAAACAATTAAAAAGTTCGGGTG
2081 L I * I F L I I G L N I K Q L K K F G W

6301 GCACAAATATNCTGTACCCTTTCTTATACACCACGGGNCTTGTCCAAATTGGGGGAGTCA
2101 H K Y X V P F L I H H G X C P N W G S H

6361 TTAATCGNGGTCAAATCTTAAATATGGGCTTTTATCAAGCTAAACACAATATGGACAAT
2121 * S X S N L K I W A F I K L N T I W T I

6421 TTAAAACTCAACCATTAATGNTG
2141 * N S T I N X

SEQ ID No. 13 DNA

SEQ ID No. 14 PRT

Lactobacillus reuteri strain ML1

1 ATCGATAATCAAATTGTTTATTTTGATATAAAGGAGATTAAAAATGGAAATAAAGAAACAT
1 I D N Q I V Y F D I K E I K M E I K K H
RBS start

61 TTTAAGTTGTATAAAAGTGGTAAACAATGGGTGACAGCGGCTGTTGCTACTGTTGCCGTT
21 F K L Y K S G K Q W V T A A V A T V A V

121 TCAACCGCGCTTCTTTACGGGGGAGTTGCACATGCTGATCAACAAGTTCAGTCTTCCACA
41 S T A L L Y G G V A H A D Q Q V Q S S T

181 ACTCAAGACCAAACCTTCTACTGTAAATACTAATACTACTAAAACAATAGCTGCAGATACT
61 T Q D Q T S T V N T N T T K T I A A D T

241 AATGCTGATCAGCCAGCTCAAACAGCTGATAAAAATCAAGCAGCATCAAATGACACTACT
81 N A D Q P A Q T A D K N Q A A S N D T T

301 AACCAAAGTAAAACTGATAGTACTTCAACAACCTGTTAAGAATCTTACTTCTACACCAGTT
101 N Q S K T D S T S T T V K N L T S T P V

361 TCTACTTTGCCATCAACTGATAATGAAAAACAAAATCAAATTATAATAAGCATGATAAT
121 S T L P S T D N E K Q N Q N Y N K H D N

421 GGAAACTATGGGAATATTGATACTGCTTACTTTAGCAATAATCAATTGCATGTTTCAGGA
141 G N Y G N I D T A Y F S N N Q L H V S G

481 TGGAATGCAACGAATGCATCTCAAGGAACAAACAGTCGGCAAATTATTGTGCGTGATATC
161 W N A T N A S Q G T N S R Q I I V R D I

541 ACAACCAATAATGAATTAGGTCGTACTGATGTAACAAACAATGTTGCGCGCCAGACGTT
181 T T N N E L G R T D V T N N V A R P D V

601 AAGAATGTTTATAATGTTTATAACGCTGATAATTCTGGATTGATATTAATGTCAATATT
201 K N V H N V Y N A D N S G F D I N V N I

661 GAATTTAGCAAGATGAAAGATTATCGGGATTCAATTGAAATTGTTAGTCGATACAGTGGA
221 E F S K M K D Y R D S I E I V S R Y S G

721 AACGGTAAATCTATTGACTGGTGGTCCCAACCGATCACTTTTGACAAAAACAATTATGCT
241 N G K S I D W W S Q P I T F D K N N Y A

781 TATCTTGATACATTTGAAGTGAAAAATGGCGAATTACATGCAACCGGATGGAATGCTACT
261 Y L D T F E V K N G E L H A T G W N A T

841 AATAGTGCAATTAACATAATCACCATTTTGTAAATTTTATTTGATCAAACGAATGGTAAG
281 N S A I N Y N H H F V I L F D Q T N G K

901 GAAGTAGCACGACAAGAAGTTCGTGAAGGCCAATCACGCCAGATGTTGCTAAGGTATAT
301 E V A R Q E V R E G Q S R P D V A K V Y

961 CCACAAGTAGTTGGTGCTGACAACCTCCGGCTTTGATGTGACATTTAATATCGGTAATTTA
321 P Q V V G A D N S G F D V T F N I G N L

1021 GATTATACTCACCAGTACCAAGTTCTTAGTCGTTACAGCAATTCTGATAATGGCGAAGGC
341 D Y T H Q Y Q V L S R Y S N S D N G E G

1081 GATAATGTTACCTACTGGTTTAATCCACAATCCATTGCTCCTGCTAATCAAAGTAACCAG
361 D N V T Y W F N P Q S I A P A N Q S N Q

1141 GGTTATCTAGACTCATTTGATATTAGTAAAAATGGTGAAGTAACAGTGACCGGATGGAAT
381 G Y L D S F D I S K N G E V T V T G W N

1201 GCTACTGACTTGTGAGAATTACAAAATAACCATTATGTAATTCTATTTGATCAGACAGCA
401 A T D L S E L Q N N H Y V I L F D Q T A

1261 GGCAACAAGTAGCATCTGCCAAGGCTGATTTAATTTACAGTCCAGATGTTGCAAAGGCT
421 G K Q V A S A K A D L I S R P D V A K A

1321 TATCCAACAGTAAAACTGCTGCAAATTCGGCTTTAAGGTAACATTTAAGGTTAATGAT
441 Y P T V K T A A N S G F K V T F K V N D

1381 TTACAACCGGGTCACCAATATAGCGTTGTAAGTCGTTTCTCTGCCGATGAAAATGGTAAT
461 L Q P G H Q Y S V V S R F S A D E N G N

1441 GGTAATGATAAGCGTCATACAGATTACTGGTTTAGTCCAGTAACATTAAACCAGAATGCT
481 G N D K R H T D Y W F S P V T L N Q N A

1501 TCAAACATTGATACTATTACAATGACATCTAATGGGTTACATATTGGCAGTTGGATGGCA
501 S N I D T I T M T S N G L H I G S W M A

1561 AGTGATAACTCAATTAATGAAACAACTCCATATGCTATTATTCTCAATAACGGTAAAGAA
521 S D N S I N E T T P Y A I I L N N G K E

1621 GTTACTCGTCAAAGATGAGTTTAACTGCCCGTCCAGATGTAGCAGCAGTATATCCTTCA
541 V T R Q K M S L T A R P D V A A V Y P S

1681 CTTTATAATAGTGCTGTTAGTGGGTTTGATACTACTATTAAATTGACTAATGATCAGTAT
561 L Y N S A V S G F D T T I K L T N D Q Y

1741 CAAGCGCTTAATGGTCAATTACAAGTATTGTTACGTTTTTCTAAAGCTGCTGATGGTAAT
581 Q A L N G Q L Q V L L R F S K A A D G N

1801 CCAAGTGGTGATAATACTGTAACGATCAATTTAGTAAAAATTATGCAACTACTGGTGGA
601 P S G D N T V T D Q F S K N Y A T T G G

1861 AACTTTGATTATGTAAAAGTAAATGGTAATCAAGTTGAATTTAGTGGTTGGCATGCAACT
621 N F D Y V K V N G N Q V E F S G W H A T

1921 AACCAATCAAATGATAAAGATTACAATGGATTATTGTTTTAGTTAATGGTAAAGAAGTA
641 N Q S N D K D S Q W I I V L V N G K E V

1981 AAGCGTCAATTAGTTAATGATACTAAAGAGGGGGCTGCTGGCTTCAACCGAAACGATGTC
661 K R Q L V N D T K E G A A G F N R N D V

2041 TACAAAGTAAATCCAGCTATTGAAAACAGTTCTATGTCTGGATTCCAAGGCATTATTACT
681 Y K V N P A I E N S S M S G F Q G I I T

2101 TTACCAGTAACAGTTAAGAATGAGAATGTTTCAGATTGTCCATCGTTTTAGTAATGATGCA
701 L P V T V K N E N V Q I V H R F S N D A

2161 AAGACAGGTGAAGGTAGCCATGTTGATTTCTGGTCAGAAGTAATGCCAGTTAAGGATAGT
721 K T _ G E G S H V D F W S E V M P V K D S

2221 TTCCAAAAGGGTAATGGTCCGCTTAAGCAATTTGGCTTACAAACTATTAATGGTCATCAA
741 F Q K G N G P L K Q F G L Q T I N G H Q

2281 TATTATATTGACCCAATGACTGGCCAACCTCGCAAGAACTTCCTATTACAAAATGGTAAT
761 Y Y I D P M T G Q P R K N F L L Q N G N

2341 GACTGGCTTTTATTTTGATAATGAAACTGGTGAGGGAACTAATGCGTTAAAGAGGCAATTT
781 D W L Y F D N E T G E G T N A L K R Q F

2401 GACGGAGGAACGATTTCTGCTGATAGTCAGTATAGAAAGGGTAATGAAGCTTATGGTTAT
801 D G G T I S A D S Q Y R K G N E A Y G Y

2461 GACAATAAGAGCATTGAAAATGTTGATGGCTTTTAAACAGCTGATACTTGGTACCGACCA
821 D N K S I E N V D G F L T A D T W Y R P

2521 AAACAAATTTTAAAATGGACCACCTGGACAGATTCTAAAGAAACAGATATGCGACCGCTC
841 K Q I L K W T T W T D S K E T D M R P L

2581 TTAATGGTTTGGTGGCCAAATACTGTAACCCAAGCATATTACCTTAACTACATGAAACAA
861 L M V W W P N T V T Q A Y Y L N Y M K Q

2641 CATGGAAACTTATTACCAGCTAATCTTCCATTCTTTAATTCTGATGCAGATCCATTAGAA
881 H G N L L P A N L P F F N S D A D P L E

2701 TTAAATTATTATGCAGAAATTGTTTCAGCAAAATATTGAAAAGAAGATTAGTCAAACCTGGT
901 L N Y Y A E I V Q Q N I E K K I S Q T G

2761 AATACTGACTGGTTGCGAACTTTGATGCACGAATTTGTATCTAATAATACAATGTGGAAT
921 N T D W L R T L M H E F V S N N T M W N

2821 AAGAATAGTGAAAATGAAGACTTTGGTGGGTTGCAATTACAAGGTGGTTTTCTAAAGTAC
941 K N S E N E D F G G L Q L Q G G F L K Y

2881 GTTAATAGTGATAAGACACCTAATGCTAATTCTAATTGGCGTATTATGGGTAGGCAGCCA
961 V N S D K T P N A N S N W R I M G R Q P

2941 GCTAATATTGACGGAAATGGGCCAATTGGATCAGAATTCTTATTAGCTAATGACGTTGAT
981 A N I D G N G P I G S E F L L A N D V D

3001 AATTCTAATCCAGTTGTTCAAGCTGAACAGTTAAATTGGCTACATTACTTATTGAATTTT
1001 N S N P V V Q A E Q L N W L H Y L L N F

3061 GGAACTATTACTGCAAATGATCCTGATGCTAATTTTGATAGCATTTCGTGTTGATGCTGTT
1021 G T I T A N D P D A N F D S I R V D A V

3121 GACAATGTAGATGCCGATTTATTAGATATAGCTGGTGATTACTTTAATGCAGTATATCAT
1041 D N V D A D L L D I A G D Y F N A V Y H

3181 TCTCAAAGTAATGATAAAATTGCTAATGCTCATATTAATATTCTTGAGGATTGGGGTGGC
1061 S Q S N D K I A N A H I N I L E D W G G

3241 CAAGATCCGTATTATACGCAAAGCATCGGAACCTCAATTATCGATGGATTATAATTTTC
1081 Q D P Y Y T Q S I G T P Q L S M D Y N F

3301 TCAACTATAAGAAGTGTGTTAGCATCTAACACTGCATCAATGACTGATATTATTAAGAAT
1101 S T I R S V L A S N T A S M T D I I K N

3361 TCATTGGTAAATCGGAGCTTAGATAATGCTGAAAACGTATCAATTCCTAATTACTCATTT
1121 S L V N R S L D N A E N V S I P N Y S F

3421 ATCCGTGCACATGATAATGGTTTCAAGATGATATTAAGCGTGCAATTTTCAGATGTAAAT
1141 I R A H D N G S Q D D I K R A I S D V N

3481 AATTTACCATATGGTTTGAAGTTTAACTTTGAGCAAGAGCAAAAGGGGATTGAAGCATAC
1161 N L P Y G S K F N F E Q E Q K G I E A Y

3541 ATTGCAGATCAAAGTAATGTTAATAAGAAGTGAATAATTATAATATTCCATCTTCATAT
1181 I A D Q S N V N K K W N N Y N I P S S Y

3601 GCTATTATGTTGACTAATAAGGATACCGTTCCTCGTGTATATTATGGTGATTATTTACT
1201 A I M L T N K D T V P R V Y Y G D L F T

3661 GATGGTGGTCAGTATATGGCACAAACAACGCGTTATTATCCTGCACTTACAAGTCTTTTA
1221 D G G Q Y M A Q T T R Y Y P A L T S L L

3721 AAGGCACGTATTAAGTATGTAGCTGGTGGACAAACAATGTCTGTGCGATAAGAATAATATT
1241 K A R I K Y V A G G Q T M S V D K N N I

3781 TTGACTAGTGTTCGCTTTGGTAAAGGTGCGATGAATCCTACTGATATGGGTGATAGTTTA
1261 L T S V R F G K G A M N P T D M G D S L

3841 ACTAGAACATCTGGTGTGGGGTAGTTATAAGTAATAATGATAAATTATTATTAAGCTCA
1281 T R T S G V G V V I S N N D K L L L S S

3901 AATGATAAAGTTGTATTACACATGGGTGCTGCACATAAGAATCAGAAATTTAAAGCAGTC
1301 N D K V V L H M G A A H K N Q K F K A V

3961 TTACTAACTACTAATGATGGTATTTCAGAGTTTTAATGATGACAATGCGCCTGTTGCATAT
1321 L L T T N D G I Q S F N D D N A P V A Y

4021 ACTGATGCTAATGGTGACTTGGTCCTTTCTGGTAAAGATATTACGACTGATGGTGTAATT
1341 T D A N G D L V L S G K D I T T D G V I

4081 CAACATAATACTGCTGTTAAGGGCTATGCTAATGCTGATGTAAAGGTTATCTTGCAGTA
1361 Q H N T A V K G Y A N A D V K G Y L A V

4141 TGGGTTCCAGTAGGTGCCAGTGATACAACAGGATATTAGAACAGCACCATCAGGGGTACAA
1381 W V P V G A S V Q Q D I R T A P S G V Q

4201 AGTGATGGAAAGTCTGTTTATCATTCAAATGCAGCTCTGGATTCAAATATTATTTTGA
1401 S D G K S V Y H S N A A L D S N I I F E

4261 GGATTCTCTAACTTTGTATATTGGCCGACAAATAATTCTGAGCGTGCAAATGTAAAAATC
1421 G F S N F V Y W P T N N S E R A N V K I

4321 GCTCAGAATACTGACTTATTTAAGGAGTTGGGTATTACTTCATTTGAATTAGCTCCACAG
1441 A Q N T D L F K E L G I T S F E L A P Q

4381 TATAATTCAAGTAAGGATGGCACATTCCTTGATTCTCAGATTGATAATGGATATGCATTT
1461 Y N S S K D G T F L D S Q I D N G Y A F

4441 ACTGATCGCTATGATCTAGGTATGAGCATTCCAAATAAGTATGGTAGCGATACTGATCTA
1481 T D R Y D L G M S I P N K Y G S D T D L

4501 AGGAATGCTATTAAAGCCTTACATAAGGCCGAATTCAAGCAATGGCTGATTGGGTTTCCT
1501 R N A I K A L H K A G I Q A M A D W V P

4561 GATCAAATTTATAATTTACCAGGTAAAGAAGTTGTTACTGCTACTCGTGTGGACGAACGT
1521 D Q I Y N L P G K E V V T A T R V D E R

4621 GGAAATGATTGGAATGTAGCTCAGATTAAGGATTCACCTTTATGTTGCTAATACAATTGGT
1541 G N D W N V A Q I K D S L Y V A N T I G

4681 GGTGGAAAGTATCAAGAGCAATATGGTGGAGCTTTCCTTGATCAATTACAAAAGCAATAT
1561 G G K Y Q E Q Y G G A F L D Q L Q K Q Y

4741 CCACAAATCTTTGAACGTAAACAACCTTCAACTGGTGTAGCAATTGACCCAAGTACTAAG
1581 P Q I F E R K Q P S T G V A I D P S T K

4801 ATTAAACAGTGGTCTGCTAAATACTTTAATGGGACAAATATTTTACATCGTGGTGCAGGG
1601 I K Q W S A K Y F N G T N I L H R G A G

4861 TATGTATTAAGAGATAACGGTGGTAACTACTTTAGCCTTGGAATAGTAATAATAACAG
1621 Y V L R D N G G N Y F S L G N S N N K Q

4921 TTATTACCAAATCAATTATCAGGTAAGGCTGAAAATGGCTTTGTTGATGTTAATGGGAAT
1641 L L P N Q L S G K A E N G F V D V N G N

4981 ACTAAATACTTTACATCAACCGGAATTCCTGTACGGATGCATTTGTTCAAGACAGTGTA
1661 T K Y F T S T G I P V T D A F V Q D S V

5041 GGTAAGTGGTACTATATTGATAAAAATGGTAATATGCTTAAAAATACCGGTTTTGTAGAT
1681 G N W Y Y I D K N G N M L K N T G F V D

5101 ATTACGCGAAATGGTCAGACAGGTACGTATCTATTCTTAAATAACGGTATCTCATTCCGA
1701 I T R N G Q T G T Y L F L N N G I S F R

5161 TCAGGATTAGTTAAAATTGGTAATGATACTTATTACTTTGACGGTAATGGAAAAATGGTT
1721 S G L V K I G N D T Y Y F D G N G K M V

5221 CGTGGCCAATCTATTAGTGATGGTACGATGAATTATACTCTTGATAAGGATGGTAAATTA
1741 R G Q S I S D G T M N Y T L D K D G K L

5281 GTTGGCTTGATTATGATCCAAGTAGTCAGAATCCACATCCAATTACTCAACAGGATTTA
1761 V G L Y Y D P S S Q N P H P I T Q Q D L

5341 AGTGGTACTAATAAGTAGTTTATTAAAAATCACCAATAGAAGTTGTCTCTACATCAAATG
1781 S G T N K * F I K N H Q * K L S L H Q M

5401 GTGTTGATATGAAAATATAATACTTTATACCATTAAATTGGTCTAGTAAGAATCATCCTC
1801 V L I * K Y N T L Y H * I G L V R I I L

5461 ACGGATGGTTCTTTTTAGTTTCGCCGTTTGTAATAAAGTTAGAAAAAATAAAAAGCCA
1821 T D G S F * F R R L * N * V R K N K K P

5521 TTTGTGATAGACTTTTGAGTATCCCTAATCAAAAGAAAGGCAATCACAAATGACCTATAA
1841 F V I D F * V S L I K R K A I T N D L *

5581 ACATCTTACCACACGCGAATTAACCTCTCATAGCTGATTTTTGGTATCAAGGCACTAAAGC
1861 T S Y H T R I N S H S * F L V S R H * S

5641 TTATCGGGCTGCTAAATACTTCAACGTAGTCAAGAAACCATCTATCGTGTATTATCGTTT
1881 L S G C * I T S T * S R N H L S C L S F

5701 CCTCAATAACGGTAAAACCATCGACCAATATCTTCAGACTTATCAGCGACATAAACGTG
1901 P Q * R * N H R P I S S D L S A T * T S

5761 TTGTGGTCGGAAGCAGACCCAACTGCCAACTATCGAGGTTAACTATATCCATGCGCAAAT
1921 L W S E A D P T A N Y R G * L Y P C A N

5821 CAAGGCTGGTTGGACTCCTGATACTATTATTGGTCGTGATGAGCACCCGATTAGCTGCAG
1941 Q G W L D S * Y Y Y W S * * A P D * L Q

5881 ATACTAATGCTGATCAGCCAGCTCAAACAGCTGATAAAAATCAAGCAGCATCAAATGACA
1961 I L M L I S Q L K Q L I K I K Q H Q M T

5941 CTACTAACCAAAGTAAAACTGATAGTACTTCAACAACTGGTAAGAATCTTACTTCTACAC
1981 L L T K V K L I V L Q Q L V R I L L L H
6001 CAGTTTTCTACTTTGGCATCAACTGATAATGGAAAACAAAATCAAATTATAATAAGCAT
2001 Q F S T L A S T D N G K Q N Q N Y N K H
6061 GATAT
2021 D

SEQ ID No. 15 DNA

SEQ ID No. 16 PRT

Lactobacillus reuteri strain ML1 (ML4)

1 AATATTGATGGTTACTTAAGTTATACTGGTTGGTATCGTCCTTATGGAACGAGTCAAGAT
1 N I D G Y L S Y T G W Y R P Y G T S Q D
61 GGTAAAACATGGTACGAAACAACCTGCAATGGATTGGCGTCCATTACTGATGTATATTTGG
21 G K T W Y E T T A M D W R P L L M Y I W
121 CCAAGCAAAGATGTTCAAGCACAATTTATTAAGTATTTTGTTAATAATGGTTATGAGAAT
41 P S K D V Q A Q F I K Y F V N N G Y E N
181 GCTAATTATGGACTTACAGAGTCCTCTGTTGCTTCCTTTAGCAAGGATACTAATGCTAAT
61 A N Y G L T E S S V A S F S K D T N A N
241 CTCCTCGATGTAACCTGCACAAAATCTTCGTTATGTAATTGAGCAAAGTATTGCAGCCAAT
81 L L D V T A Q N L R Y V I E Q S I A A N
301 AAAGGGACAAGTAAGTTAGCAAATGATATTAATAGTTTTGCTGCAACGGTTCCTGAATTA
101 K G T S K L A N D I N S F A A T V P E L
361 TCTGCATCATCTGAATTATCATTGCAAAGCATGCCAACTATCGACCAGATGAAAGTGGA
121 S A S S E L S L Q S M P N Y R P D E S G
421 ACTGTTGATAGTGATCAAGTCATTTTGTTAATAATAATTCAAAGGATCCCCGTAAAGGG
141 T V D S D Q V I F V N N N S K D P R K G
481 AACACTGGTTATGCGGACAGCAACTATCGCTTAATGAACAGGACGATTAATAATCAGGCC
161 N T G Y A D S N Y R L M N R T I N N Q A
541 GGAAATAATAATAGTGATAACAGTCCAGAACTCCTTGTTGGTAATGATATTGATAATTCA
181 G N N N S D N S P E L L V G N D I D N S
601 AACCAGTAGTACAAGCTGAAAATCTTAATTGGGAATACTTTTTACTAAATTATGGTAAG
201 N P V V Q A E N L N W E Y F L L N Y G K
661 TTAATGGGGTATAATCCAGACGGTAATTTTGATGGCTTCCGAGTTGATGCTGCTGATAAT
221 L M G Y N P D G N F D G F R V D A A D N
721 ATTGATGCAGATGTCTTAGATCAAATGGGTCAATTAATGAACGACATGTATCATACAAAG
241 I D A D V L D Q M G Q L M N D M Y H T K
781 GGAAATCCTCAAATGCAAATGATCATTGAGTTATAATGAAGGTTATCATTCTGGGGCT
261 G N P Q N A N D H L S Y N E G Y H S G A
841 GCACAAATGCTAAATGAAAAGGGTAATCCTCAATTGTACATGGATTTCAGGCGAATTCAT
281 A Q M L N E K G N P Q L Y M D S G E F Y
901 ACCCTTGAGAATGTTCTCGGACGTGCAAATAACCGTGATAGTATCGGTAATTTAATTACT
301 T L E N V L G R A N N R D S I G N L I T

961 AATAGTGTGTTAATCGGCAAAATGATACAACAGAGAATGAAGCTACGCCAAACTGGTCA
321 N S V V N R Q N D T T E N E A T P N W S

1021 TTTGTAACCTAACCATGATCAACGAAAGAATTTGATTAATAGATTAATTATTAAGGGTCAT
341 F V T N H D Q R K N L I N R L I I K G H

1081 CCTAACATTCCGGATATTATGGGTTTACAAAGCTGAATATGCAATCAAGCATGG
361 P N I P D I M G S A Y K A E Y A N Q A W

1141 CAAGAATTCTACGCTGATCAGAAAAAGACTAATAAACAATATGATCAATATAATGTTCCG
381 Q E F Y A D Q K K T N K Q Y D Q Y N V P

1201 GCTCAGTATGCAATTCTTTTGAGCAATAAAGATACGGTTCCGCAGGTTTACTATGGTGAC
401 A Q Y A I L L S N K D T V P Q V Y Y G D

1261 CTTTATAATGAACTGCTCAATACATGCAAGAGAAGTCAATTTACTATGATACAATCAGC
421 L Y N E T A Q Y M Q E K S I Y Y D T I T

1321 ACTCTTATGAAGGCCCGTAAACAATTTGTTAGTGGTGGTCAAACGATGACTAAACTTAAC
441 T L M K A R K Q F V S G G Q T M T K L N

1381 AATAATTTATTAGCTAGTGTTCGATATGGTAAGGGTGTGCTGATTCTAATAGCAATGGT
461 N N L L A S V R Y G K G V A D S N S N G

1441 ACCGATAAGCTTAGCCGAACAAGTGGGATAGCCGTCTTAGTTGGTAATGATAGTAATATG
481 T D K L S R T S G I A V L V G N D S N M

1501 GCTCAACAACTGTTGCTATTAATATGGGTCGCGCTCATGCTAACCAACAATATCGAAAT
501 A Q Q T V A I N M G R A H A N Q Q Y R N

1561 TTAATTGATACTACCGAAAAATGGCTTGACATATGATGGAGAAAATAGTGAAAATCCAGCC
521 L I D T T E N G L T Y D G E N S E N P A

1621 ATTTTGACAACTGATAGTAATGGTATCTTAAAAGTAACAGTTAAAGGATACAGTAACCCA
541 I L T T D S N G I L K V T V K G Y S N P

1681 TACGTAAGTGGTTATCTTGGTGTGTTGGGTTCCAGTAATTTCTGGTGATCAAGATGTTACT
561 Y V S G Y L G V W V P V I S G D Q D V T

1741 ACAAGTGCAAGTGATGTTGTTGCTGATAAAGAAAAGACTTTTGAATCTAATGCTGCTCTT
581 T S A S D V V A D K E K T F E S N A A L

1801 GATTCTCATATGATCTATGAAGATTTTACGCTTGTTCCAACCAGAACCAACTAATGTTGAG
601 D S H M I Y E D F S L F Q P E P T N V E

1861 AATCATGCTTACAATGTGATTGCTAAAAATGCTAATCTCTTTAATGATTTAGGCATTACT
621 N H A Y N V I A K N A N L F N D L G I T

1921 GATTTTTGGATGGCTCCTGCTTACACTCCATTTGGAATGAGTCGTTATAATGAAGGATAC
641 D F W M A P A Y T P F G M S R Y N E G Y

1981 TCAATGACGGATCGTTACAATTTAGGTACGACAGCTAATCCAACAAAGTATGGTAGTGGA
661 S M T D R Y N L G T T A N P T K Y G S G

2041 GAAGAGCTTGCAAATACAATTGCTGCATTGCATAAAGTAGGATTAAAAGTTCAAGAAGAT
681 E E L A N T I A A L H K V G L K V Q E D

2101 ATTGTTATGAATCAGATGATTGGTTTCTCTGGTCAAGAAGCAGTAACGGTTACTCGAACA
701 I V M N Q M I G F S G Q E A V T V T R T

2161 AATAATCGTGGAATGCAGATTCATGTAAATGGTCAAACATATGCAATCAAATTTACTTT
721 N N R G M Q I H V N G Q T Y A N Q I Y F

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2221 GCATATACAACCTGGTGGCGGAAATGGTCAAGAACTTATGGTGGTAAATACCTTGCCGAA
741 A Y T T G G G N G Q E T Y G G K Y L A E

2281 TTACAAAAGAACTATCCTGACCTATTTACGACCAAGGCAATTTGACAGAAAGTTGTACCT
761 L Q K N Y P D L F T T K A I S T E V V P

2341 GATCCAACCGTTTCGTATTAAT
781 D P T V R I N

SEQ ID No. 17 DNA

Lactobacillus strain LB33

1 ATGGAATTAA AAAGGCATTA CAAGATGTAC AAGGCTGGTA AAAAATGGGT TTTTGCTGCA
61 ATTGCCACAA TCTCTATAAT TGCAGGATTA AATACAGTGG CAGTGACAAC CTATGCTGCC
121 GGCAATAATG ATCCGCAGCA GACCACTACT CAAATGCAC CTAACAACAG TAACGATCCG
181 CAATCTACTA CTACGCAGAA TACTGCCAAC AACAGTAACG ATCCGCAATC TACTACTACG
241 CAGAATACTG CCAACAACAG TAATGGTCCA CAATCTACTA CTACGCAGAA TACTGCCAAC
301 AATAGTAATG GTCCACAATC TACTACTACG CAGAATACTG CCAATAACAG TAACGATCCA
361 CAATCTACTA CTACGCAGAA TACTGCCAAC AACAGTAACG ATCCGCAATC TACTACTACG
421 CAGAATACTG CCAACAATAG TAATGGTCCA CAATCTACTA CTACGCAGAA TACTGCCAAC
481 AACAGTAACG ATCCGCAATC TACTACTACG CAAAACACTG CCAACAACGG TAATGATCCA
541 CAATCTACTA CTGGAAAAGA TACAGTTAGT ATTGCAGATA TTCAAGTTAA CCAACCTGTT
601 AATCTTTTAG GAAAGCAATC AACTGTATCT AGTACTGGTT ATAATGACTC TCACATAAAA
661 AATGTCAATG GGAAAATCTA TTTTGTGGT GATAATGGTC AGGTCAAGAA AAACCTTACA
721 GCCATAATCA ATGGACAATC ACTATATTTT AATAAAACAA CTGGAGAATT GGCTTCTAAT
781 GATGTTCAAT ATGAAAATGG GTTAGTAAAA ATAAACGATG TTCATAACGC CGCTTACTCT
841 ATTGATCCAA CGGGATTAC TAATGTTAAC GGATTTTTAA CTGCTAATAG TTGGTATAGA
901 CCCAAATATA TTTACAAAGA TGGGCAAAAA TGGGTGGAAT CAACCTCTCA AGATATGCGT
961 CCCCTTTTAA TGACATGGTG GCCAGATAAA AATACTCAAG TAGCTTATTT ACAATATATG
1021 CAGAAAATGG GCATTTTACC CGCTGACGTC ACTATATCAA GTCAAACCAA TCAATCAGTT
1081 TTAACCAAAG AATCATTAT TACTCAAGCT GAAATTGAAA AACAGATTGG AGTAACAAAT
1141 GGAAACACTG ATTGGCTAAA GAAAGATATC TCTGATTTTG TAAATTCTCA ACCAAATTGG
1201 AATATAGATA GTGAAGCCAA AGGCACAGAC CATTTCAGG GGGGAGCACT TTTATATGTT
1261 AATAATAAGT TAATCCATA TCGAATTCT GATTACCGCT TGCTTAACCG AACACTTACT
1321 AATCAACAGG GGCAAGTAAA AGATACTTCT AAACAAGGCG GTTATGAAAT GTTACTTGCC
1381 AACGATGTGG ATAATTCCAA TCCAGTAGTT CAAGCGGAAC AGTTAACTG GTTATACTAC
1441 ATGATGAATA TAGGTAGCAT TACTGCCAAT GATCCACCG CAACTTTGA TGGCTATCGA
1501 GTGGACGCTG TGGACAATGT CGATGCTGAT TTATTAAATA TAGCTGCCGA TTATGCCAAA
1561 GATGCTTATA AAATAATCA AAGTGATGCT AATGCCAACA AACATTTATC AATATTAGAA
1621 GATTGGGATA ATAATGATCC GGCTTATATC AAAGCACATG GAAATAATCA GTTAACATG
1681 GATTTCACAG CACATTTAGC AATTAAATAT TCATTAAATA TGCCAGTAAG TCAACGAAGT
1741 GGGCTGGAAC CAGAGCTCAC AACCAGTTTA GTTAACAGAA CTGGTGATGA TTCTACTGAA
1801 AATGTCGCAC AGCCAAACTA TACTTTTATT AGGGCTCACG ATAGTGAAGT GCAAACAATC
1861 ATCGCACAAA TTATCAAAGA TAAATCAAC CCTAACTCTG ACGGATTAAC AGTTACTCCC
1921 GATGAAATAA GTCAGGCCTT TAAATATAT AATGCAGATG AATTAAAGAC TGATAAACAA
1981 TATACTTTTT ATAACATGCC CTCTGCCTAT ACTATTTTGC TAACCAATAA AGATACAGTA
2041 CCTCGAGTTT ATTATGGGGA TCTTTATAGT GATAATGGCA ATTATATGTC AGCCCATCTCT
2101 CCTTACTATG ATGCAATAAC TACGTTATTA AAAACACGAA TGAAATACGT ATCTGGTGGT
2161 CAAAACATGC GTATGCAATA TATGCAGGGT GATGATATGC CTGCTAATAG CTATAAGGGC
2221 GTTTTAACTT CAGTTAGATA TGGTAAGGGT GAAATGACAG CCGATGAGCA AGGTAATTCA
2281 GAAACTCGTA CTCAAGGAAT TGGGGTCATT ATAAGCAATA ATCCTAATTT AAAATTAGAC
2341 AGTAATGACC AAGTGGTATT AAATATGGGG GCGGCACATG AAAATCAAAC TTATCGCCCT
2401 GTATTACTAA CAACTAAAGA TGGATTGAAA AACTATGATT CCGATAGTTC TGTACCTCAA
2461 AATGCATTAG TTTCAACCAA CGATAAGGGA CAACTCATAT TTAAAGCTAG TTCTATTACG
2521 GGAGTAAGTA ATCCGCAGGT ATCTGGTTAT TTGTCCGTGT GGGTCCCAGT GGGGGCAAAG
2581 GATAATCAAG ATGCTCGGAC TGCAAGCAGT TCTCAGCCAT CAACTGATGG CAAAACATAT
2641 CAATCCAATG CTGCTTTAGA CTCTCAAGTT ATTTACGAAG GATTTTCTAA TTTTCAATCG
2701 ATTCCTACAA ATACAGAAGA TTTCACTAAT GTAAAAATTG CTCAAAACGC TAACCTGTTT
2761 AAGAGCTTGG GAATAACAAG TTTTGAATTA GCCCTCAAT ATCGTTCAG TAATGATAAT
2821 AGTTTTCTGG ATTCGGTTGT TCAAAATGGC TACGCATTTA CTGATCGTTA TGATATTGGG
2881 TATAATACTC CGACAAAATA TGGAACTGTT ACTCAATTGC TGGATGCATT AAGGGCTTTA
2941 CATGCCAACG GAATTCAGC GATCGATGAC TGGGTTCCCTG ACCAAATATA CAATTTACCT
3001 GGTGAGGAAA TTGTCGCAGC TCAAAGAACT AATGGATCTG GGACATATGA TCAAGATTCT

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3061 GTTATTGATG ATACATTATA TGATTCTCAC ACTGTTGGTG GTGGCGAATA TCAAGCTAAA
3121 TTTGGTGGAG CTTTTCTAAA CAAGTTAAAG CAGTTGTATC CTGATTTATT TAAAGTTAAA
3181 CAAATTTCTA CTGGTCAACC TATGAATCCT AATGAAAGAA TTACCGAGTG GTCAGCAAAG
3241 TACTTTAATG GTACAAATAT TCAAGGAAGA GGCGCTTGGT ATGTATTAAA AGACTGGGGT
3301 ACCAATCAGT ACTTTAATGT AAGTAATAAC CAGTTTGTTT CCAAACAATT CCTAGGTACA
3361 GATACTTATA CAGGCTTTAA TGTTACAAAT GAGGGAACTC AGTTTTATT C TACGAGTGGG
3421 TATAAAGCCC AGAATACCTT TATTCAGGAC GGAGACAACT GGTATTACTT TGACAATAAT
3481 GGCTATATGG TAACTGGTTT ACAGAATATA AATGGGAATA ATTACTATTT CTGCCCCAAT
3541 GGCATTGAAC TACAAGACTC TTATTTATTG AATGATGATA CCGGTAAAGA ATATTATTAT
3601 GCAAGTAATG GTAAGCAAAT CTCAAATCGT TATTATCCAG ATGCTAACGG CAATTGGAGA
3661 TATTTCTTCA ATGATGGTTC AATGGCAAGA AATGGATTAA CCACTATTGA ACAACCAGAT
3721 GGGCAAAAAG TGATCCAATA TTTTGATTCC GATGGTATTC AATTAAAGGG AAATGCCGCA
3781 AAAGATAATA ATGGTAATTT AAGATATTTT GACGGTAATA CAGGTGATAT GGTCAATTAAT
3841 TCATTTGGAG AACTTCCTGA TGGCTCTTGG TTATACCTTA ATGATAAGGG GATTGCCGTT
3901 ACTGGTAAAC AGGAAATCAA TGGTCAAACC TACTACTTTG ATGCGGATGG CAAGCAAAGTG
3961 AAGAATGATT TTAGAGAGTT GCCTGATGGT TCATGGCTTT ATCTTAATGA CAAGGGGATT
4021 GCCGTTACTG GTAAACAGGA AATCAATGGT CAAACCTACT ACTTTGATGC GGATGGCAAG
4081 CAAGTGAAGA ATGATTTTAG AGAGTTGCCT GATGGTTCAT GGCTTTATCT TAATGACAAG
4141 GGGATTGCCG TTACTGGTAA ACAGGGAATC AATGGTCAA CCTATGCAGA GGCTAAAATC
4201 ACAGCTGCCG AAAATGCTCA TCAAGCTGCC ACAGACGCTG TGAATAAAGC CCAAGCTGCT
4261 CAATCGCCTA ACACTAGTTC CTCTAGTTCT AGCGTTAGCC AAGCTACTAA ACATCAATTG
4321 GCAGTTAAAA CTGCTAAAGC TCAACTTGCT AAAACTAAGG CTCAAATTGC TAAGTATCAA
4381 AAGGCTTTGA AAAAAGCCAA AACTACAAAG GCCAAGGCTC AAGCTCGTAA AAGTTTGAAG
4441 AAGGCCGAGA CTAGTTTCAG CAAAGCTGAA CTTAATTTGG CATTATTAAA TAATAAAGCC
4501 GTAAAAGCTG CACAAACTAA GGTAAATAAG GCTAAGGCTC AAGTCACTAA ATACCAAAG
4561 GCTTTGAAGA AAGCTAAGAC TACAAAGGCT AAGACTCAAG CTCGTAAAAA TTTGAAGAAG
4621 GCCAACTCTA GTCTGACAAA AGCTCAAAAA GCATTAACTA AAGTAATTAA AACCAATATC
4681 AAGTAA

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SEQ ID No. 18 PRT

Lactobacillus strain LB33

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MELKRHYKMYKAGKKWVFAA IATISIIAGLNTVAVTTYAA
GNNDPQQTQTQONAPNNSNDP QSTTTQNTANNSNDPQSTTT
QNTANNSNGPQSTTTQNTAN NSNGPQSTTTQNTANNSNDP
QSTTTQNTANNSNDPQSTTT QNTANNSNGPQSTTTQNTAN
NSNDPQSTTTQNTANNGNDP QSTTGKDTVSIADIQVNQPV 200
NLLGKQSTVSSTGYNDSHIK NVNGKIYFVGDNQVKKNFT
AIINGQSLYFNKTTGELASN DVQYENGLVKINDVHNAAYS
IDP?GFTNVNGFLTANSWYR PKYIYKDGQKWVESTSQDMR
PLLMTWVDPKNTQVAYLQYM QKMGIPLADV TISSQTNQSV
LTKESEFITQAEIEKQIGVTN GNTDWLKKDISDFVNSQPNW 400
NIDSEAKGTDHLQGGALLYV NNKLTPYANS DYRLNRTLT
NQOQVQVQKTSKQGGYEMLLA NDVDNSNPVVQAEQLNWLYY
MMNIGSITANDPTANFDFGYR VDAVDNVDADLLNIAADYAK
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VLLTTKDG LKNYDS DSSVPQ NALVSTNDKGQLIFKASSIQ
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YNTPTKYGTVTQLLDALRAL HANGIQ AIDDWVPDIYNLP 1000
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GQKVIQYFSDSGIQLKGNAA KDNNGNLRYFDGNTGDMVIN
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QVKNDFRELPDGSWLYLNDK GIAVTGKQGINGQTYAEAKI 1400
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AVKTAKAQLAKT?AQIAKYQ KALKKAKTTKAKAQARKSLK
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SEQ ID No. 19 DNA

SEQ ID No. 20 PRT

Lactobacillus sake strain KG15

1 SASCTGBCMSTNACGTTHRRCNTAGACGTTHRACGTACTGGTTCACACAATGGATTCCGGC
1 X X X X R X X X T X X V L V H T M D S A

61 AAACATCAATGATTGCGATCTGTCCAGGTTGGGCTGCTTCACGCGTCAAACCGTACGG
21 N Y Q * L R S V Q V G L L H A S N Q Y G

121 ATCGCATTGACCACGGGTAATAATTGTAGTGC GCGACGGTTGAACCGTGACCGACTAATG
41 S H * P R V I I V V R D G * T V T D * W

181 GTGATTTTTTTCGGGCATAAAGGCGGTCAATCAAGCGCCAAAACGGCGTTGTGATTGAATA
61 * F F A A * R R S S S A K N G V V I E Y

241 CCAAGCGTTGTTTGTAACACAGTAGCGCCAACAATCGACAGTCATCGATTTTAACGTGC
81 Q A L F V N T V A P T I D S H R F * R A

301 GCCACATTACGCCGTTGCGTCACACAACGTGGGCAATAGCGCTGGTAAAGCGACTGGCAC
101 P H Y A V A S H N V G N S A G K A T G T

361 AGCTGATAAAAATAATGATAGTTACCTTGTAATTCGTGACGAATTTGTTTAAACTTAGGA
121 A D K N N D S Y L V I R D E F V * T * D
- 35

421 TGGTTCAACATCGTTAGGACCCCTTTTAAGTTTAGTCACTTATGAATCTAACTGTGTGG
141 G S T S L G P L L S L V T Y E S N C V G
- 10

481 ACTTTTTTGTTAATTTTTTTGTATTATTACAACTAGCACCACGCGTATGTGTTTTATTA
161 L F C * F F C I I T N * H H A Y V F Y *
RBS

541 ATACCACTTAATTAATAACGGGGCTTTAGCATGATTTCAAATAAAATAGTGTGAAAGGTA
181 Y H L I N N G A L A * F Q I K * C E R *
start

601 GTTTTTTATGTTAAGGAATAATTATTTTGGAGAGACTAAAACGCATTATAAATTATATAA
201 F F M L R N N Y F G E T K T H Y K L Y K

661 ATGCGGTAAGAACTGGGCTGTCATGGGGATTTTCAATTTCCGCTGGGATTAGGGATGCT
221 C G K N W A V M G I S L F P L G L G M L

721 AGTTACCAGCCAGCCAGTGTGCTGATGTGACAGCCACCAGCACCTCAAGCAGTGCAGT
241 V T S Q P V S A D V T A T S T S S S A V

781 GAGGACCGATGCAATCAGTGCAAGTAGTAGCAGTGCAGCAAAGGCTGAAACGGCTGCGAT
261 R T D A I S A S S S S A A K A E T A A I

841 CACTACTGCAGGTGTTGCAAATGCTGATTCACAAACATCAGCAGAAGTAACCGCTGACTC
281 T T A G V A N A D S Q T S A E V T A D S

901 TACTTCTACCAGCCAAGTGGTAATAATAATTCCAATAATCAAAATAATACAGCACAGCC
301 T S T S Q V V T N N S N N Q N N T A Q P

961 AGCCGGTCAAGAAGCAGCCCCGGTATCAGAGGACACATCATCTGATGATAGTGAGAGAAC
321 A G Q E A A P V S E D T S S D D S E R T

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2281 TCAGGGTGTATGGTAACGGGTAAGCAACGTGTGCACCAAGATCAGTATTTCTTCCTGCC
761 Q G V M V T G K Q R V H Q D Q Y F F L P

2341 AAATGGTATTGCTTTGACAGATGCTTTTCGTACAAACTGCTGATGGTCAACGTCAGTACTA
781 N G I A L T D A F V Q T A D G Q R Q Y Y

2401 TGATAAAACAGGTCGTCTGGTCATTAATCAATATGTGACTGACCACCAAGCGAATGCGTT
801 D K T G R L V I N Q Y V T D H Q A N A F

2461 CCGGGTTGATGCAGACGGTAACGTTGTCCGCAATCAAGCTTTGACTGTTGACGGCCATGA
821 R V D A D G N V V R N Q A L T V D G H E

2521 ACAATATTTCCGGCACAACGGTGTCCAAGCGAAAGCAGTGCTCATTGAACTGACGATAA
841 Q Y F G T N G V Q A K A V L I R T D D N

2581 TCAGGCGCGCTACTACGAAGCCAATAGTGGTAATCTCGTGAAGCAACAGTTTATTCTTGA
861 Q A R Y Y E A N S G N L V K Q Q F I L D

2641 TACAGATGGACATTGGTTGTACGCGGATGCTGCAGGTGACTTGGCACGCGGACAAATTAC
881 T D G H W L Y A D A A G D L A R G Q I T

2701 AATTGGCCAAGACACGTTGTATTTTGATGATAATAATCACCAGGTAAAAGATGATTTTCGT
901 I G Q D T L Y F D D N N H Q V K D D F V

2761 CTATGATACTAACGGTGTGCATTATTTTAATGGCACAACAGGCGCTGAAATCAAACAAGA
921 Y D T N G V H Y F N G T T G A E I K Q D

2821 TTACGCGTTTCATGATGGCAAATGGTACTATTTTGATGATTTGGGACGAATGGTAACCGG
941 Y A F H D G K W Y Y F D D L G R M V T G

2881 CTTGCAGCGTATTAATGGTGAGTATCGCTATTTTGATGCTAATGGTGTGCAACTAAAGGG
961 L Q R I N G E Y R Y F D A N G V Q L K G

2941 CGGTACCGTGACCGATCCACTAACGCACCAAACGTACACTTTTGATGCGAAAACCTGGTGC
981 G T V T D P L T H Q T Y T F D A K T G A

3001 TGGTACGTTGGTGACGATTTAACTGAATAATGGACTAGAAAAGACGATCTTGTATCGTCT
1001 G T L V T I * L N N G L E K T I L Y R L

3061 TTTTLAGTTTCGATAACTAAATAAGTGCTCATTTTGCATTAGGACTCAGAATTAGCGGG
1021 F * F R * L N K C S F L H * D S E L A G

3121 CGCGCAAGCGTCTTTTCGTGTTAACTTATTAGTAATTAATATTTTGAGGAGTCTGTTAT
1041 A Q A S F R V K L I S N * Y F E E S V I

3181 ATGGCAACAATTTTAGTTGTAGATGATGAACCGTCATTGGTGACGCTACTGTCATACAAC
1061 W Q Q F * L * M M N R H W * R Y C H T T

3241 CTGACTAAATCAGGCTTCGAGGTCGTGACTGCTACCTCCGGTGACGAGGCACGAAATCAG
1081 * L N Q A S R S * L L P P V T R H E I S

3301 CTGGCAAATCATCCTATTGATTTGATGCTGCTAGGTGTCATGTTGCCTGGTAAGAGTGGC
1101 W Q I I L L I * C C * V S C C L V R V A

3361 GTTGACTTAACACGAGAACTACGAGGCGAACAAGAATCGTATTCCAATTATTATGATTACC
1121 L T * H E N Y E A N R I V F Q L L * L P

3421 GCCTTGGATGACGAAGTTGACAAGATTT
1141 P W M T K L T R F